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Atherosclerosis and Inflammation the Path from Pathogenesis to Treatment: Review of the Current State of the Issue (Part 2)

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ABSTRACT

Numerous studies addressing the fundamental aspects of atherosclerosis emphasize the importance of systematically organizing the accumulated data. The second part of this lecture provides an analysis of the critical mechanisms involved in the development of atherosclerosis. This analysis includes a discussion on the roles of inflammasomes, hemodynamic disorders within the vascular wall, vasa vasorum pathology, endothelial cell dysfunction, matrix metalloproteinases, and the Notch and Wnt signaling pathways in the process of atherogenesis. Additionally, it explores the specific characteristics of the pathogenesis of vascular calcification associated with atherosclerosis. A dedicated section thoroughly reviews contemporary pharmacotherapeutic strategies for managing atherogenic dyslipidemia. A comprehensive analysis of current concepts regarding the pathogenesis of atherosclerosis, along with promising approaches to drug therapy, will facilitate the identification of future research directions within the field of lipidology. This endeavor has the potential to elevate preventive cardiology to a new standard.

Keywords: atherosclerosis, inflammation, inflammasome, atheroma, PCSK9 inhibitors

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Атеросклероз и воспаление – путь от патогенеза к терапии: обзор современного состояния проблемы (часть 2)

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РЕЗЮМЕ

Достижения многочисленных исследований в изучении фундаментальных аспектов атеросклероза диктуют необходимость систематизации накопленных данных. Во второй части лекции представлен анализ роли ключевых механизмов реализации воспалительного процесса в развитии атеросклероза. Рассмотрена роль инфламмасомы, нарушений гемодинамики в сосудистой стенке, патологии *vasa vasorum*, дисфункции эндотелиоцитов, матриксных металлопротеиназ, сигнальных путей Notch и Wnt в атерогенезе, а также ассоциированные с атеросклерозом особенности патогенеза кальцификации сосудов.

Отдельным разделом представлен обзор современных фармакотерапевтических подходов к лечению атерогенной дислипидемии. Комплексный анализ современных представлений о патогенезе атеросклероза и перспективных методов лекарственной терапии позволит обозначить дальнейшие направления исследований в липидологии и вывести возможности профилактической кардиологии на потенциально новый уровень.

Ключевые слова: атеросклероз, воспаление, инфламмазома, атерома, ингибиторы PCSK9

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INTRODUCTION

Atherosclerosis is one of the primary challenges in preventive cardiology, which has traditionally received significant attention in the development of national programs for the primary and secondary prevention of atherosclerosis-associated cardiovascular diseases (aCVD) and cardiac rehabilitation programs [1-4].

According to data from the multicenter study ESSE-RF, which included respondents aged 25-64 years from 13 regions of the Russian Federation (RF), the prevalence of hypercholesterolemia (total cholesterol (TC) in the blood ≥ 5.0 mmol/L) averaged $58.40 \pm 0.34\%$. This indicates an extremely high frequency of atherogenic dyslipidemia within the study population [5]. In the United States, data from the National Health and Nutrition Examination Survey revealed that levels of TC over 200 mg/dL and low-density lipoprotein cholesterol (LDL) ≥ 130 mg/dL were found in 32.8% and 36.2% of examined individuals, respectively [6].

According to the multicenter, cross-sectional, observational study EURIKA (European Study on Cardiovascular Risk Prevention and Management in Usual Daily Practice), which included data from 12 countries (Austria, Belgium, Germany, France, Greece, Turkey, and others, including Russia) with a final sample size of 7,641 patients, the proportion of individuals with atherogenic dyslipidemia was over 20% [7]. The EURIKA population comprised European patients aged at least 50 years who had at least one risk factor for cardiovascular disease (CVD) but no history of CVD in their medical records. Additionally, the STEPs 2021 study reported that the proportion of individuals with atherogenic dyslipidemia (based on all lipidogram indicators) among the population of the Islamic Republic of Iran was 81.0% [8].

A cross-sectional study conducted as part of the China-PEACE project involved 2,660,666 individuals aged 35 to 75 years from all provinces of the People's Republic of China between 2014 and 2019. Among those examined, the prevalence of atherogenic dyslipidemia was found to be 33.8% [9].

These findings indicate that atherogenic dyslipidemia is a global problem, as evidenced by the prevalence rates of lipid metabolism disorders observed across diverse populations with varying national dietary habits. Consequently, studying the pathogenesis of atherosclerosis and developing new therapeutic methods aimed at normalizing lipid

metabolism and stabilizing inflammatory status are critically important. The role of inflammation in the development of ASCVD is well established and underscores the urgency of this research.

Currently, atherosclerosis is perceived by the scientific community as an inflammatory disease of the arteries that triggers the mechanisms of vascular aging and damage to target organs [10, 11]. Given this fact, the study of atherogenesis problems from the standpoint of inflammatory theory is a relevant fundamental direction with direct access to real clinical practice [12-16].

In the second part of this lecture, attention will be directed towards examining the clinically relevant aspects of inflammation pathogenesis in the context of atherosclerosis development. Furthermore, a summary of therapeutic methodologies, grounded in the latest progressions in clinical lipidology, will be presented.

The Role of Inflammasome in Atherogenesis

In the context of the leading role of inflammation in the pathogenesis of atherosclerosis, it is worth emphasizing the role of the inflammasome, since this intracellular multiprotein complex is known to play a crucial role in the relationship between lipid metabolism and low-grade inflammation of the vascular wall [17]. Cholesterol crystals and oxidized lipoproteins activate monocytes and macrophages, generating an inflammatory response followed by the production of proinflammatory interleukins (IL) - IL-1 β and IL-18. Oxidized LDL is recognized by CD36 receptors on recruited monocytes, which leads to activation of the NLRP3 inflammasome [18]. In lipopolysaccharide (LPS)-treated monocytes, saturated fatty acids can induce the release of IL-1 β , which is not observed with unsaturated fatty acids [19]. Like monocytes, endothelial cells also demonstrate NLRP1 activation after stimulation with plasma containing high levels of triacylglycerols and VLDL [20]. In addition to lipid metabolism disorders, other mechanisms are involved in triggering atherogenesis-associated inflammation.

Hypoxia and hypoxia-associated signaling through hypoxia-inducible factor (HIF)-1 α in atherosclerotic plaques enhance NLRP3 expression in macrophages and slow the degradation of proIL-1 β [21]. Hemodynamically induced shear stress increases the expression of sterol regulatory element-binding protein 2 (SREBP2) via mechanotransduction, triggering a new wave of atherogenesis. In this context, elevated NLRP3 expression in endothelial

cells plays an crucial role in maintaining aberrant lipid metabolism [22]. The development of dysfunctional autophagy in atherosclerotic plaques is also significant in the process of atherogenesis, as evidenced by the increased expression of autophagy markers ATG13 and LC3 in aortic endothelial cells. Notably, in mice lacking the ATG5 protein which is essential for autophagy, there is an increase in inflammatory activity and plaque size. These findings underscore the importance of autophagy in the pathogenesis of ASCVD [23]. In mice fed a high-cholesterol diet, hematopoietic deletion of NLRP3, ASC, or IL-1 α /IL-1 β resulted in reduced atherogenesis and lower levels of IL-18 [24]. Furthermore, pharmacological inhibition of NLRP3 with colchicine increases the number of smooth muscle cells (SMCs) and collagen within the atherosclerotic plaque, promoting its transition to a more stable phenotype [25].

Vascular Shear Stress and Atherosclerosis

Under normal conditions, uniform laminar blood flow acting on the intima of the arteries induces the secretion of nitric oxide (NO). In turn, NO released under physiological conditions regulates the tone of the vascular wall and helps maintain the anti-inflammatory and antithrombotic properties of the endothelium. It is well established that the formation and progression of atheroma occurs focally, primarily around bifurcations or at the points where lateral branches depart from the artery, that is, in areas characterized by uneven (turbulent) blood flow [26]. This nature of the blood flow creates low wall shear stress (WSS), which induces vascular inflammation and contributes to the development of atherosclerosis. WSS refers to the tangential force of mechanical friction exerted by flowing blood, acting longitudinally on the endothelium surface of the arterial wall [27].

Specific endothelial biomechanical receptors within the endothelial glycocalyx detect mechanical stimuli and differentiate between laminar and turbulent types of blood flow, converting WSS into biochemical signals [28]. Consequently, endothelial dysfunction induced by WSS is closely linked to inflammation and lipid metabolism disturbances in the vascular wall, thereby promoting the progression of atherosclerosis. It is worth noting that, although atherogenesis initially occurs in regions of the arterial wall exposed to low WSS, areas of high WSS that develop around growing atherosclerotic plaques are associated with the formation of an unstable plaque phenotype

[29]. As WSS increases, the functioning of the mechanoreceptor KLK10 diminishes, which mediates the transformation of the normal transcriptome signature of arteries into an emergency response profile [30]. Inflammatory changes within the plaque lead to hypoxia, initiating neovascularization from the adventitial vasa vasorum, which contributes to increased plaque vulnerability [31]. In discussing the vasa vasorum, it is important to highlight the theory that atherosclerosis may initiate specifically from these microvessels within the vascular wall of the arteries [32]. The microvascular network of the vasa vasorum (including arterial, venous, and lymphatic vessels of varying calibers) serves as a crucial anatomical and functional structure that meets the metabolic needs of the adventitia and perivascular adipose tissue, as well as the outer part of the medial layer of large arteries [33]. Dysregulation of blood flow in the vasa vasorum is implicated in the pathogenesis of atherosclerosis, as evidenced by the presence of multiple neuroimmune cardiovascular interfaces (NICIs) in the outer layers of atherosclerotic arteries. These interfaces are characterized by axon terminals located near the SMC media and macrophages in perivascular adipose tissue [34]. Numerous newly formed vasa vasorum are abundant in lipid-rich plaques and express elevated levels of cell adhesion molecules, such as ICAM-1 and VCAM-1. This expression facilitates an excessive influx of immune cells and is associated with plaque instability [35].

Although the concept of initial vasa vasorum pathology in the initiation of atherogenic changes currently has several gaps, their role in atherogenesis is extremely important, both within the framework of the “outside-in” concept and in the classical approaches to study. During vascular wall inflammation, vascular endothelial (VE) cadherin is phosphorylated by Src kinase 3 at the intercellular junctions of the endothelium. Concurrently, dephosphorylation of VE cadherin by VE protein tyrosine phosphatase (VE-PTP) prevents its internalization and stabilizes the adhesive junctions between endothelial cells [36, 37].

Additionally, the dissociation of VE-PTP from VE cadherin leads to leukocyte diapedesis and increased vascular permeability in vivo, as demonstrated in a model induced by vascular endothelial growth factor (VEGF) and endotoxin [38]. It is known that lymphocyte binding to the adhesion molecule VCAM-1, along with the stimulation of endothelial cells by VEGF, triggers a common signaling cascade

that includes Ras-associated botulinum toxin substrate C3, NADPH oxidase, reactive oxygen species, and proline-rich tyrosine kinase 2 [39, 40]. However, the molecular mechanisms regulating the kinetics of the interaction between VE-PTP and VE-cadherin remain largely unexplored. Signaling protein 2 containing the CUB-EGF domain (SCUBE2) ensures the integrity of the vascular wall by recruiting VE-PTP to dephosphorylate VE-cadherin. This process promotes the stabilization of endothelial adherens junctions and preserves the barrier function of the intima [41]. Studies involving genetic overexpression and pharmacological induction of SCUBE2 further support the concept that therapeutic regulation of SCUBE2 may be beneficial for stabilizing the vascular bed [42].

Inflammation also stimulates the development of dystrophic calcification in the necrotic lesion of atherosclerotic plaques as a healing response to the inflammatory activation of macrophages [43]. The death of macrophages and SMCs releases vesicles that serve as “nucleation sites” for the deposition of hydroxyapatite crystals. Their aggregation leads to the formation of microcalcifications with diameters of less than 50 μm , which can penetrate the fibrous cap of the plaque [44, 45]. Microcalcifications significantly contribute to the instability of atherosclerotic plaques; furthermore, they induce mechanical stress within the fibrous capsule, generating new inflammatory impulses within the plaque [46]. It is also important to note that ectopic deposition of calcium hydroxyapatite salts occurs long before the onset of atherocalcinosis.

In atherosclerotic inflammation, various cell types, including vascular SMCs, resident pericytes, circulating stem cells, and adventitial cells, differentiate into osteoblastic cells, leading to vascular calcification [47]. For example, SMCs lose part of their contractile phenotype, as evidenced by downregulation of α -smooth muscle actin (α -SMA) and SM-22 expression, followed by abnormal upregulation of genes involved in osteogenesis, such as Runt-related transcription factor 2 (Runx2), osteopontin, osteocalcin, etc. [48, 49]. Vascular calcification is initiated by matrix vesicles produced by osteoblast-like cells that serve as deposition sites for hydroxyapatite crystals [50]. Meanwhile, the overexpression of matrix metalloproteinase MMP-9 leads to the degradation of elastin, which in turn promotes the transition of SMCs from a contractile to a producing phenotype [51].

The Role of Inflammation in Plaque Destabilization

Atherosclerotic plaques are primarily composed of extracellular matrix (ECM), which includes collagen, elastin, proteoglycans, and glycosaminoglycans synthesized by SMCs in the arterial wall [52]. Under conditions of atherogenic inflammation, cytokines such as IL-1 β and tumor necrosis factor α (TNF- α) induce the secretion of metalloproteinases, particularly MMP-1, MMP-8, MMP-9, MMP-12, and MMP-13, by macrophages under the regulation of microRNA [53-55].

MMPs catalyze the destruction of interstitial collagen, leading to thinning and weakening of the fibrous capsule, which contributes to plaque instability [56]. In addition, the stability of the fibrous capsule is influenced by the cross-linking of collagen fibers, a process mediated by the enzyme lysyl oxidase (LOX), which is expressed by endothelial cells [57]. Endothelial dysfunction and the phenotypic transition of SMCs are associated with a decrease in LOX activity, resulting in abnormal collagen cross-linking. This weakens the fibrous capsule and increases the presence of soluble collagen forms that are subject to MMP-mediated degradation [58].

In unstable atherosclerotic plaques, the activity of MMP-7 and MMP-9 is increased, and tissue expression of MMP-2 and MMP-9 raises alongside a decrease in the expression of type IV collagen [59]. Among the three types of unstable atheromas, lipid-type plaques exhibit the highest tissue expression of MMP-9 compared to dystrophic-necrotic and inflammatory-erosive types, while type IV collagen expression is predominant in dystrophic-necrotic atherosclerotic plaques. In addition to MMPs, an 8-fold significant increase in APOE gene expression ($p < 0.001$) was observed in unstable atherosclerotic plaques of the dystrophic-necrotic type. In contrast, stable atherosclerotic plaques showed an 8-fold statistically significant increase in LDLR and APOB gene expression ($p < 0.001$) [60].

Interestingly, the level of adiponectin in an atherosclerotic plaque is directly proportional to serum levels of HDL-C, while secretin levels are inversely proportional. Furthermore, the glucagon levels in conditionally intact intima are 2.1 times lower than those in fragments with stable atherosclerotic plaque; it has also been established that secretin levels are directly associated with plaque stability [61].

In recent decades, more and more attention has been paid by researchers to such a phenomenon as atherosclerotic plaque erosion. Plaques that have undergone superficial erosion demonstrate less lipid accumulation, a less pronounced necrotic core, a moderate number of inflammatory cells, and an intact fibrous capsule [62]. Thrombi formed as a result of superficial erosions are white and rich in platelets, while thrombi associated with plaque rupture are red (rich in fibrin and erythrocytes) [63].

Parallels between Notch and Wnt Signaling Pathways and Atherosclerosis

Notch is a cellular signaling pathway that mediates intercellular communication and is involved in the regulation of homeostasis [64]. The Notch cascade protects against endothelial dysfunction induced by pro-inflammatory cytokines and regulates the phenotypic transition of cells [65]. Increasing evidence suggests that Notch plays a crucial role in signaling related to changes in WSS [66].

Activation of the Notch pathway creates an anti-inflammatory, anti-atherogenic environment that helps maintain endothelial integrity, including the preservation of adherens junctions between endothelial cells [67]. Additionally, Notch is a key signaling cascade for regulating the structure and function of SMCs. Expression of Notch receptors 2 and 3, as well as the primary ligand Jagged1, has been observed in SMCs [68]. Mutations in Notch 2 and 3 can lead to defects in SMC development, providing a strong evidence for the involvement of Notch signaling in regulating vascular differentiation during angiogenesis [69]. Furthermore, Jagged1-Notch3 signaling mediated through nidogen-2 is essential for maintaining the contractile phenotype of SMCs in vitro and in vivo [70].

Wnt is a multitarget signaling cascade characterized by three main intracellular signaling pathways: the canonical pathway (Wnt/ β -catenin), the non-canonical Wnt/PCP pathway (which regulates cytoskeletal dynamics through the activation of JNK (C-Jun N-terminal kinase) by small G proteins), and the Wnt/ Ca^{2+} -dependent pathway [71]. In addition to its roles in cell proliferation and differentiation, the Wnt pathway is also involved in regulating lipid metabolism [72]. The stabilization of β -catenin via Wnt signaling, along with the activation of fatty acid synthesis via Akt/mTOR signaling, plays a central role in lipid metabolism in steatotic liver [73]. An inverse relationship has been demonstrated between

Wnt activation and the severity of atherosclerosis. Specifically, activation of the Wnt pathway following lipid depletion enhances the IL-4 response in macrophages via the PGE2/STAT3 axis. Dickkopf-2 (DKK2), a negative regulator of Wnt/ β -catenin signaling, is implicated in macrophage activation during atherosclerosis [74].

Knockdown of DKK2 significantly reduces the expression of genes associated with the polarization of macrophages toward the pro-inflammatory M1 phenotype while increasing the level of polarization markers associated with the anti-inflammatory M2 phenotype. This knockdown also significantly attenuates the formation of foam cells [75].

The Role of MicroRNA in the Pathogenesis of Atherosclerosis

The role of microRNA in atherosclerosis is multifaceted. For example, miR-520c-3p protects endothelial cells from damage and stabilizes endothelial function by regulating key aspects of pathogenesis, such as cell proliferation, apoptosis, and endothelial cell adhesion [76]. Moreover, miR-181a-5h, miR-181a-3p, and miR-250b modulate the severity of chronic low-grade inflammation in the vascular wall by suppressing the expression of the nuclear factor NF- κ B, thereby slowing the progression of stromal-vascular dystrophic changes [77]. Conversely, miR-488 [78] and miR-183-5p [79] exhibit proatherogenic effects by stimulating functional reorganization of SMCs and exacerbating inflammatory infiltration in the vascular wall. MicroRNAs also demonstrate a dual effect on macrophages. Thus, miR-10a, miR-210, and miR-383 stabilize mitochondrial metabolism and the redox status of cells, leading to a reduction in apoptosis and necroptosis [80]. Notably, miR-181a-3p/5p and miR-155-5p have pronounced atheroma-stabilizing effects [81]. However, high levels of miR-155 correlate with NLRP3 activation via ERK1/2 kinase [82]. In addition, miR-216a exhibits proatherogenic potential by enhancing inflammation through the Smad3/NF- κ B cascade [83].

A Look at Lipid-Lowering Therapy through the Prism of the Inflammatory Theory of Atherogenesis

In parallel with the active study of the molecular mechanisms of atherogenesis, the drug arsenal of lipid-lowering therapy is expanding, which increases the capabilities of modern cardiology.

The basic drugs of lipid-lowering therapy are traditionally considered to be HMG-CoA reductase

inhibitors – statins (in particular, rosuvastatin, pitavastatin and atorvastatin) both without and in combination with ezetimibe - a selective inhibitor of cholesterol absorption targeting the sterol transporter Neimann-Pick-like1 (NPC1L1) [84]. This combination is considered generally accepted and complies with the recommendations of both the Russian and European Cardiology Societies.

In the context of this lecture, it is important to focus on the anti-inflammatory potential of statins. Analyzing the mechanism of action of statins reveals that part of their pleiotropic effects can be attributed to the blockade of the mevalonate pathway of cholesterol synthesis, which reduces the levels of isoprenoid intermediates such as farnesyl pyrophosphate and geranyl-geranyl pyrophosphate. A decrease in these levels changes the prenylation of proteins, influencing the effects of statins on autophagy and inflammation [85]. Moreover, statins can suppress the adhesion and migration of inflammatory cells by reducing the expression of the integrin dimer CD11, the immunoglobulin superfamily protein VCAM-1, and leukocyte functional antigen-1 (LFA-1). They also decrease the expression of monocyte chemotactic protein-1 (MCP-1) and interleukin-8 (IL-8) [86].

Another anti-inflammatory mechanism of statins is their ability to reduce the levels of interferon γ (INF- γ), oxidized LDL (oxLDL), and serum apoA-I [87, 88]. Several potential mechanisms through which statins exert their anti-inflammatory effects via Toll-like receptor (TLR) signaling pathways have also been identified: inhibition of the prenylation of regulatory proteins, direct or indirect inhibition of NF- κ B and MyD88/NF- κ B axis, and activation of antioxidant response elements (ARE) [89]. In addition, statins can reduce signaling mediated by transforming growth factor TGF- β 1 in T lymphocytes, suppress oxLDL-induced maturation of human dendritic cells, impair T lymphocyte activation, and stimulate the pool of regulatory T lymphocytes [90]. Further studies are needed to elucidate the complete molecular mechanisms and multifaceted anti-inflammatory potential of statins. At the same time, several issues persist regarding statin use, particularly their side effects, such as statin-induced myopathy and hyperglycemia. Other concerns include partial and complete resistance to statins, the presence of residual cardiovascular risk, and elevated levels of triglyceride-rich lipoproteins, despite achieving target levels of total cholesterol, LDL cholesterol, and triacylglycerols

[91-97]. In light of these challenges, new drugs aimed at normalizing cholesterol metabolism are currently being actively developed and introduced into clinical practice. Among the extensive list of lipid-lowering agents, the most promising include

1) PCSK9-modifying agents

Proprotein convertase subtilisin-kexin type 9 (PCSK9) inhibitors, particularly evolocumab and alirocumab, are innovative drugs that are actively utilized in modern clinical practice [98-100]. The pivotal studies demonstrating the lipid-lowering potential of evolocumab and alirocumab are FOURIER [101] and ODYSSEY-OUTCOMES [102] trials. According to a meta-analysis of 41 randomized clinical trials, which included a cumulative sample of 76,304 patients (49,086 received evolocumab and 27,218 received alirocumab), PCSK9 inhibitors significantly reduce the risk of myocardial infarction, coronary artery restenosis, and ischemic stroke. Furthermore, these agents are well-tolerated and considered safe drugs while effectively lowering LDL cholesterol levels [103]. In addition to their significant beneficial effects on lipid metabolism and the reduction of major adverse cardiovascular outcomes (MACE) [104], PCSK9 inhibitors also demonstrate significant anti-inflammatory effects. A study from the European Collaborative Project on Inflammation and Remodeling of the Vascular Wall in Intravascular Ultrasound (ATHEROREMO-IVUS) demonstrated that serum PCSK9 levels are associated with increased absolute inflammatory plaque volume and necrotic core size [105]. A clear correlation was also observed between serum PCSK9 levels and the concentrations of pro-inflammatory cytokines, including IL-6, IL-1 β , TNF- α , macrophage colony-stimulating factor (M-CSF), and high-sensitivity C-reactive protein (hs-CRP) [106]. It has been established that PCSK9 enhances the infiltration of inflammatory monocytes into the vessel wall due to the interaction of PCSK9-LDLR (less pronounced with LRP5) with plaques. This interaction directly contributes to plaque destabilization [107]. PCSK9 itself induces inflammation and exacerbates atherosclerosis independently of the LDL receptor. Research has shown that PCSK9 worsens atherosclerosis in mice with a knockout of the LDL receptor gene. Adenylate cyclase-associated protein 1 (CAP1) serves as the primary transducer for mediating the inflammatory actions of PCSK9, including the induction of cytokines, Toll-like receptor 4, scavenger receptors, and the lectin-type oxidized low-density lipoprotein receptor

1 (LOX-1) [108]. Key mediators of this inflammatory cascade include spleen tyrosine kinase (Syk) and protein kinase C delta (PKC δ), which are activated following the formation of the PCSK9-CAP1 complex [109]. In human peripheral blood mononuclear cells, it has been established that PCSK9 levels positively correlate with the phosphorylation of Syk, PKC δ and p65 [110]. Thus, the anti-inflammatory effect of PCSK9 inhibition is evident and holds significant clinical relevance. In discussing drug approaches targeting PCSK9, it is important to highlight inclisiran, a drug based on small interfering RNA (siRNA) [111, 112]. Inclisiran is a double-stranded modified siRNA linked to N-acetylgalactosamine (GalNAc), which acts as a ligand for the asialoglycoprotein receptor expressed by hepatocytes. The drug specifically binds to the matrix RNA transcribing the sequence of the gene encoding PCSK9 [113]. By disrupting the translation of PCSK9 through mRNA cleavage, inclisiran effectively reduces its production. The ORION study series [114] provides robust evidence regarding its hypolipidemic potential, supported by meta-analyses [115, 116] that confirm its clinical efficacy in achieving target lipidogram indicators and reducing adverse cardiovascular outcomes. It is worth noting that some studies within the ORION series are still ongoing today.

2) Lipoprotein (a) inhibitors

Lipoprotein (a) or Lp(a), is an independent factor contributing to both overall and residual risk of CVD [117, 118]. Individuals with elevated Lp(a) levels (>125 nmol/L; >50 mg/dL) exhibit increased activity of arterial inflammation, characterized by endothelial activation due to oxidized phospholipids carried by Lp(a). This process leads to the recruitment of circulating monocytes, resulting in heightened secretion of chemoattractants and pro-inflammatory cytokines, increased expression of adhesion molecules, and enhanced leukocyte migration through the vascular wall [119]. Unfortunately, lifestyle modifications have minimal impact on Lp(a) levels; therefore, extracorporeal therapies, such as namely lipoprotein apheresis may be necessary. This approach is supported by latest American Heart Association consensus on LP(a) apheresis published in 2024 [120]. Lp(a) particles can cross the endothelial barrier, persist in the arterial wall, and promote the development of atherosclerotic plaques [121]. The oxidized phospholipids carried by Lp(a) can trigger macrophage apoptosis and contribute to the “instability” of atheromas [122]. Additionally,

Lp(a) promotes inflammation within the arterial wall by increasing monocyte extravasation and endothelial activation [123].

These effects are mediated through adhesion molecules such as ICAM-1 and are associated with an increase in the activity of the enzyme 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase (PFKFB)-3 induced by Lp(a) [124]. The development of drugs targeting high Lp(a) levels represents an innovative approach to lipid-lowering therapy, as elevated Lp(a) levels are a strong and independent risk factor for ASCVD. As of 2024, several drugs have emerged in this category: pelacarsen [125], olpasiran [126], zerlasiran [127], lepodisiran [128], and muvalaplin [129]. Notably, clinical trials involving these agents have generated great interest within the scientific community, particularly studies such as OCEAN(a)-DOSE [130], KRAKEN [131], ALPACAR [132], among others.

3) Antisense oligonucleotides

Volanesorsen and olezarsen are antisense oligonucleotides targeting apolipoprotein C3 (APOC3) mRNA and are currently under active investigation for the treatment of familial chylomicronemia syndrome [133]. Volanesorsen blocks the synthesis of apolipoprotein C3 in the nucleus of hepatocytes by inhibiting APOC3 mRNA. Two main clinical trials have been conducted with volanesorsen: APPROACH [134] and its open-label extension (OLE) [135], as well as the COMPASS trial [136]. Olezarsen represents an advancement over volanesorsen, as it is conjugated to N-acetylgalactosamine, an aminosaccharide that exhibits a strong binding affinity for the asialoglycoprotein type 1 receptor, thereby enhancing its targeting to hepatocytes [137]. Evidence supporting the efficacy of olezarsen comes from a double-blind, placebo-controlled study [138], which demonstrated that olezarsen reduces levels of apolipoprotein C3, triacylglycerols, and atherogenic lipoproteins in patients with moderate hypertriacylglycerolemia who are at high risk or have established cardiovascular disease.

4) Bempedoic acid

Bempedoic acid is a long-chain tetramethyl-substituted ketodiac acid characterized by a linear molecule structure. It belongs to the family of “rogue” fatty acids [139].

As a hypolipidemic agent, bempedoic acid functions as an inhibitor of the enzyme ATP-citrate lyase, which catalyzes one of the key reactions in cholesterol synthesis [140]. It is the first drug in its

class to act by inhibiting adenosine triphosphate citrate lyase [141]. A significant aspect of bempedoic acid's mechanism of action is that its active metabolite is formed exclusively in the liver, which minimizes the risk of muscle-related adverse reactions [142]. The safety and efficacy of long-term use of bempedoic acid have been evaluated in the CLEAR (Cholesterol Lowering via BEMPedoic Acid, an ACL-inhibiting Regimen) program, which encompasses four phase 3 studies: CLEAR Tranquility [143], CLEAR Harmony [144], CLEAR Wisdom [145], and CLEAR Serenity [146].

Bempedoic acid promotes the activation of LDL receptor expression, leading to lower LDL cholesterol levels, attenuation of atherogenesis, reduction in hepatocyte lipid levels and body weight, and improvement in glycemic control [147, 148]. In this regard, both genetic inhibition of ATP-citrate lyase (ACLY) in hepatocytes and pharmacological inhibition with bempedoic acid suppress fatty acid and cholesterol synthesis while enhancing fatty acid oxidation without increasing circulating triacylglycerol levels. Moreover, studies conducted on murine and human hepatic stellate cells have demonstrated that bempedoic acid also inhibits liver fibrosis by targeting pathways involved in collagen formation [149].

5) Evinacumab

Evinacumab is a monoclonal antibody that targets angiopoietin-associated peptide 3 (ANGPTL3), a circulating protein secreted by the liver that regulates the hydrolysis of very low-density lipoprotein (VLDL) triglycerides. This drug is typically used for the treatment of refractory homozygous familial hypercholesterolemia [150].

6) Lomitapide

Lomitapide lowers cholesterol levels by inhibiting microsomal triacylglycerol transfer protein (MTP) [151]. MTP is involved in loading triacylglycerols onto apolipoprotein B100, which is essential for VLDL assembly. After being secreted by hepatocytes, VLDL is converted to LDL. By blocking VLDL assembly, lomitapide reduces both VLDL release and VLDL-mediated triacylglycerol secretion, resulting in lower plasma LDL concentrations [152]. Lomitapide has been approved by the FDA and EMA for the treatment of adult patients with homozygous familial hypercholesterolemia as an adjunct to a low-fat diet and other lipid-lowering therapies, with or without LDL apheresis [151]. Despite the impressive therapeutic potential of new drugs, their use is

limited due to the lack of large-scale double-blind randomized studies, insufficient clinical experience, and high costs. Consequently, they are considered reserve therapies and are prescribed in cases where target lipid profile indicators are not achieved with the maximum tolerated dose of statins combined with ezetimibe and/or when there is complete intolerance to statins [153, 154].

7) Colchicine

In the context of trends in contemporary cardiology, it is worthwhile to highlight the role of colchicine in the treatment of atherosclerosis. Colchicine is a significant medication whose mechanism of action is linked to its effects on cellular structure and function. This drug exhibits a biphasic effect on microtubules; at low concentrations, it inhibits microtubules growth, while at high concentrations, it promotes their depolarization [155]. Colchicine inhibits tubulin polymerization, disrupting the cellular cytoskeleton and leading to impairment of various intracellular processes, including mitosis, intracellular transport, and phagocytosis [156]. In addition, colchicine inhibits chemotaxis and the adhesion of neutrophils to inflamed endothelium, including indirectly through alterations in the expression of VE-selectin on endothelial cells [157]. Colchicine also inhibits L-selectin expression, preventing neutrophil recruitment, and affects neutrophil function by limiting their extravasation. Furthermore, colchicine normalizes macrophage activity and inflammasome functioning [158]. Beyond its effect on neutrophils, colchicine exhibits antithrombotic activity by reducing leukocyte-platelet aggregation (including both monocytes and neutrophils) as well as lowering levels of surface markers associated with platelet activity, such as P-selectin and PAC-1 (activated GP IIb/IIIa) [159].

Thus, the diverse effects of colchicine, including modulation of the cell cytoskeleton, anti-inflammatory properties, and antithrombotic activity, determine its high clinical significance in reducing both overall and residual cardiovascular risk in atherosclerosis [160]. There is a substantial body of evidence supporting the use of colchicine in atherosclerosis; notable studies include COLCOT (COLchicine Cardiovascular Outcomes) [161], LoDoCo (Low Dose Colchicine) [162], COVERT-MI (Colchicine for Left Ventricular Infarct Size Reduction in Acute Myocardial Infarction) [163], and CONVINCENCE (Colchicine for prevention of Vascular Inflammation in Non-CardioEmbolic Stroke) [164].

8) Biologically active compounds in contemporary lipidology

In parallel with conventional drug therapy, the role of various biologically active substances with hypolipidemic activity is being actively studied. Notable examples include chitosan, ursolic acid, nattokinase, spermidine, taurine, grape and pomegranate seed extracts, as well as many other naturally derived compounds that are positioned as atheroprotective and hypolipidemic substances [165, 166]. This topic is traditionally considered controversial. Unfortunately, the available data on the effectiveness and safety of these compounds are limited, difficult to compare, and sometimes even contradictory. Nonetheless, this does not exclude their potential benefits, which have been supported by large placebo-controlled, double-blind randomized studies. For example, the COSMOS (COcoa Supplements and Multivitamin Outcomes Study) study demonstrated a 27% reduction in cardiovascular mortality rates associated with cocoa flavonoids [167]. Additionally, a network meta-analysis encompassing 131 studies with a total sample size of 13,062 patients compared the effectiveness of various dietary supplements such as artichoke, berberine, bergamot, garlic, green tea extract, plant sterols/stanols, policosanols, red yeast rice, silymarin, and spirulina. This analysis found that bergamot and red yeast rice extracts exhibited the most significant atheroprotective effect [168]. It is important to note that in the vast majority of cases, while the positive effects of these compounds are statistically significant compared to placebo groups, they are not comparable to those of statins. The interpretation of data from existing studies is further complicated by the high variability in the biological properties of natural raw materials. These properties can depend on factors such as the life cycle conditions of the producing organisms and the conditions under which they are harvested, processed, and stored. Therefore, caution should be exercised when interpreting these findings. However, the significance of these results should not be underestimated; they should be considered in clinical practice, particularly, when developing personalized dietary interventions that align with clearly defined treatment goals.

CONCLUSION

Our understanding of atherosclerosis has evolved significantly beyond the concept of a mere lipid metabolism disorder. Contemporary research highlights the pivotal role of inflammation throughout

the entire atherosclerotic process. Notably, both innate and adaptive immune responses are activated in atherosclerosis, initiating inflammatory reactions that occur both locally and systemically, manifesting as chronic low-grade inflammation. Consequently, circulating cytokines not only serve as indicators of heightened cardiovascular risk but also actively contribute to the progression and destabilization of atherosclerotic plaques. Understanding the role of inflammation in the pathogenesis of atherosclerosis presents significant clinical implications. The pursuit of identifying a molecular signature of the inflammatory cascade in atherosclerotic cardiovascular disease (aCVD) may facilitate the development of targeted anti-inflammatory strategies in the future. When combined with personalized medicine approaches, this advancement could significantly enhance the capabilities of preventive cardiology.

REFERENCES

1. Boytsov S.A., Pogosova N.V., Ansheles A.A., Badtieva V.A., Balakhonova T.V., Barbarash O.L. et al. Cardiovascular prevention 2022. Russian national guidelines. *Rossiyskiy kardiologicheskii zhurnal - Russian Journal of Cardiology*. 2023;28(5):5452. (In Russ.). DOI:10.15829/1560-4071-2023-5452.
2. Pogosova N.V., Oganov R.G., Boytsov S.A., Ausheva A.K., Sokolova O.Yu., Kursakov A.A. et al. Secondary prevention in patients with coronary artery disease in Russia and Europe: results from the Russian part of the EUROASPIRE V survey. *Kardiovaskulyarnaya terapiya i profilaktika - Cardiovascular Therapy and Prevention*. 2020;19(6):2739. (In Russ.). DOI: 10.15829/1728-8800-2020-2739
3. Pogosova N.V., Boytsov S.A. Preventive Cardiology 2024: State of the Problem and Perspectives of Development. *Cardiology*. 2024;64(1):4-13. (In Russ.). DOI:10.18087/cardio.2024.1.n2636.
4. Pogosova N.V. The Importance of Cardiac Rehabilitation in the Era of Modern Cardiovascular Disease Treatment. *Cardiology*. 2022;62(4):3-11. (In Russ.). DOI:10.18087/cardio.2022.4.n2015.
5. Metelskaya V.A., Shalnova S.A., Deev A.D., Perova N.V., Gomyranova N.V., Litinskaya O.A. et al. Analysis of the Prevalence of Indicators Characterizing Atherogenicity of Lipoprotein Spectrum Among the Population of the Russian Federation (Based on the ESSE-RF Study). *Preventive Medicine*. 2016;19(1):15-23. (In Russ.). DOI:10.17116/profmed201619115-23.
6. Zheutlin A.R., Harris B.R.E., Stulberg E.L. Hyperlipidemia-Attributed Deaths in the U.S. in 2018-2021. *Am. J. Prev. Med.* 2024;66(6):1075-1077. DOI: 10.1016/j.amepre.2024.02.014.
7. Halcox J.P., Banegas J.R., Roy C., Dallongeville J., De Backer G., Guallar E. et al. Prevalence and treatment of atherogenic dyslipidemia in the primary prevention of cardiovascular disease in Europe: EURIKA, a cross-sectional observational study. *BMC Cardiovasc. Disord.* 2017;17(1):160. DOI: 10.1186/s12872-017-0591-5.

8. Khanali J., Ghasemi E., Rashidi M.M., Ahmadi N., Ghamari S.H., Azangou-Khyavy M. et al. Prevalence of plasma lipid abnormalities and associated risk factors among Iranian adults based on the findings from STEPs survey 2021. *Sci. Rep.* 2023;13(1):15499. DOI: 10.1038/s41598-023-42341-5.
9. Lu Y., Zhang H., Lu J., Ding Q., Li X., Wang X. et al. China patient-centered evaluative assessment of cardiac events million persons project collaborative group. Prevalence of dyslipidemia and availability of lipid-lowering medications among primary health care settings in China. *JAMA Netw. Open.* 2021;4(9):e2127573. DOI: 10.1001/jamanetworkopen.2021.27573.
10. Majee S., Banerjee A. Suppression of inflammatory macrophages reduces atherosclerosis. *J. Physiol.* 2024;602(16):3867–3869. DOI: 10.1113/JP287013.
11. Ali I., Zhang H., Zaidi S.A.A., Zhou G. Understanding the intricacies of cellular senescence in atherosclerosis: Mechanisms and therapeutic implications. *Ageing Res. Rev.* 2024;96:102273. DOI: 10.1016/j.arr.2024.102273.
12. Aldana-Bitar J., Golub I.S., Moore J., Krishnan S., Verghese D., Manubolu V.S. et al. Colchicine and plaque: A focus on atherosclerosis imaging. *Prog. Cardiovasc. Dis.* 2024;84:68–75. DOI: 10.1016/j.pcad.2024.02.010.
13. Avagimyan A., Fogacci F., Pogoseva N., Kakturskiy L., Jndoyan Z., Faggiano A. et al. Methotrexate & rheumatoid arthritis associated atherosclerosis: A narrative review of multidisciplinary approach for risk modification by the international board of experts. *Curr. Probl. Cardiol.* 2024;49(2):102230. DOI: 10.1016/j.cpcardiol.2023.102230.
14. Abdelmaseih R., Alsamman M.M., Faluk M., Hasan S.M. Cardiovascular Outcomes With Anti-Inflammatory Therapies: Review of Literature. *Curr. Probl. Cardiol.* 2022;47(6):100840. DOI: 10.1016/j.cpcardiol.2021.100840.
15. Poznyak A.V., Bharadwaj D., Prasad G., Grechko A.V., Sazonova M.A., Orekhov A.N. Anti-inflammatory therapy for atherosclerosis: focusing on cytokines. *Int. J. Mol. Sci.* 2021;22(13):7061. DOI: 10.3390/ijms22137061.
16. Ridker P.M. The time to initiate anti-inflammatory therapy for patients with chronic coronary atherosclerosis has arrived. *Circulation.* 2023;148(14):1071–1073. DOI: 10.1161/CIRCULATIONAHA.123.066510.
17. Theofilis P., Oikonomou E., Chasikidis C., Tsioufis K., Tousoulis D. Inflammasomes in atherosclerosis-from pathophysiology to treatment. *Pharmaceuticals (Basel).* 2023;16(9):1211. DOI: 10.3390/ph16091211.
18. Ionov A.Yu., Kuznetsova E.A., Kindalyova O.G., Kryuchkova I.V., Poplavskaya E.E., Avagimyan A.A. Clinical significance of endocrine disorders in the development of early vascular aging in males with abdominal obesity and concomitant arterial hypertension: An observational cohort study. *Kuban Scientific Medical Bulletin.* 2024;31(1):74–87. DOI: 10.25207/1608-6228-2024-31-1-74-87.
19. L'homme L., Esser N., Riva L., Scheen A., Paquot N., Piette J. et al. Unsaturated fatty acids prevent activation of NLRP3 inflammasome in human monocytes/macrophages. *J. Lipid. Res.* 2013;54(11):2998–3008. DOI: 10.1194/jlr.M037861.
20. Bleda S., de Haro J., Varela C., Ferruelo A., Acin F. Elevated levels of triglycerides and vldl-cholesterol provoke activation of nlrp1 inflammasome in endothelial cells. *International Journal of Cardiology.* 2016;220:52–55. DOI: 10.1016/j.ij-card.2016.06.193.
21. Folco E.J., Sukhova G.K., Quillard T., Libby P. Moderate hypoxia potentiates interleukin-1 β production in activated human macrophages. *Circ. Res.* 2014;115(10):875–883. DOI: 10.1161/CIRCRESAHA.115.304437.
22. Xiao H., Lu M., Lin T.Y., Chen Z., Chen G., Wang W.C. et al. Sterol regulatory element binding protein 2 activation of NLRP3 inflammasome in endothelium mediates hemodynamic-induced atherosclerosis susceptibility. *Circulation.* 2013;128(6):632–642. DOI: 10.1161/CIRCULATIONAHA.113.002714.
23. Lin L., Zhang M.X., Zhang L., Zhang D., Li C., Li Y.L. Autophagy, pyroptosis, and ferroptosis: new regulatory mechanisms for atherosclerosis. *Front. Cell Dev. Biol.* 2022;9:809955. DOI: 10.3389/fcell.2021.809955.
24. Silvis M.J.M., Demkes E.J., Fiolet A.T.L., Dekker M., Bosch L., van Hout G.P.J. et al. Immunomodulation of the NLRP3 inflammasome in atherosclerosis, coronary artery disease, and acute myocardial infarction. *J. Cardiovasc. Transl. Res.* 2021;14(1):23–34. DOI: 10.1007/s12265-020-10049-w.
25. Martínez G.J., Celermajor D.S., Patel S. The NLRP3 inflammasome and the emerging role of colchicine to inhibit atherosclerosis-associated inflammation. *Atherosclerosis.* 2018;269:262–271. DOI: 10.1016/j.atherosclerosis.2017.12.027.
26. Wang X., Shen Y., Shang M., Liu X., Munn L.L. Endothelial mechanobiology in atherosclerosis. *Cardiovasc. Res.* 2023;119(8):1656–1675. DOI: 10.1093/cvr/cvad076.
27. Qaisar S., Brodsky L.D., Barth R.F., Leier C., Buja L.M., Yildiz V. et al. An unexpected paradox: wall shear stress in the aorta is less in patients with severe atherosclerosis regardless of obesity. *Cardiovasc. Pathol.* 2021;51:107313. DOI: 10.1016/j.carpath.2020.107313.
28. Roux E., Bougaran P., Dufourcq P., Couffignal T. Fluid shear stress sensing by the endothelial layer. *Front. Physiol.* 2020;11:861. DOI: 10.3389/fphys.2020.00861.
29. Sergin I., Evans T.D., Bhattacharya S., Razani B. Hypoxia in plaque macrophages: a new danger signal for interleukin-1 β activation? *Circ. Res.* 2014;115(10):817–820. DOI: 10.1161/CIRCRESAHA.114.305197.
30. Zhou Z., Korteland S., Tardajos-Ayllon B., Wu J., Chambers E., Weninck J. et al. Shear stress is uncoupled from atheroprotective KLK10 in atherosclerotic plaques. *Atherosclerosis.* 2024;398:118622. DOI: 10.1016/j.atherosclerosis.2024.118622.
31. Shirai K., Hitsumoto T., Sato S., Takahashi M., Saiki A., Nagayama D. et al. The Process of Plaque Rupture: The role of vasa vasorum and medial smooth muscle contraction monitored by the cardio-ankle vascular index. *J. Clin. Med.* 2023;12(23):7436. DOI: 10.3390/jcm12237436.
32. Tinajero M.G., Gotlieb A.I. Recent developments in vascular adventitial pathobiology: the dynamic adventitia as a complex regulator of vascular disease. *Am. J. Pathol.* 2020;190(3):520–534. DOI: 10.1016/j.ajpath.2019.10.021.
33. Elmarasi M., Elmakaty I., Elsayed B., Elsayed A., Zein J., Boudaka A., Eid A. Phenotypic switching of vascular smooth muscle cells in atherosclerosis, hypertension, and aortic dissection. *J. Cell Physiol.* 2024;239(4):e31200. DOI: 10.1002/jcp.31200.

34. Mohanta S.K., Peng L., Li Y., Lu S., Sun T., Carnevale L. et al. Neuroimmune cardiovascular interfaces control atherosclerosis. *Nature*. 2022;605(7908):152–159. DOI: 10.1038/s41586-022-04673-6.
35. Sluiter T.J., Buul J.D., Huveneers S., Quax P.H., de Vries M.R. Endothelial barrier function and leukocyte transmigration in atherosclerosis. *Biomedicines*. 2021;9(4):328. DOI: 10.3390/biomedicines9040328.
36. Juettnner V.V., Kruse K., Dan A., Vu V.H., Khan Y., Le J. et al. VE-PTP stabilizes VE-cadherin junctions and the endothelial barrier via a phosphatase-independent mechanism. *J. Cell Biol.* 2019;218(5):1725–1742. DOI: 10.1083/jcb.201807210.
37. Nawroth R., Poell G., Ranft A., Klop S., Samulowitz U., Fachinger G. et al. VE-PTP and VE-cadherin ectodomains interact to facilitate regulation of phosphorylation and cell contacts. *EMBO J.* 2002;21(18):4885–4895. DOI: 10.1093/emboj/cdf497.
38. Broermann A., Winderlich M., Block H., Frye M., Rossaint J., Zarbock A. et al. Dissociation of VE-PTP from VE-cadherin is required for leukocyte extravasation and for VEGF-induced vascular permeability *in vivo*. *J. Exp. Med.* 2011;208(12):2393–2401. DOI: 10.1084/jem.20110525.
39. Zhang S., Zhang Q., Lu Y., Chen J., Liu J., Li Z. et al. Roles of Integrin in Cardiovascular Diseases: From Basic Research to Clinical Implications. *Int. J. Mol. Sci.* 2024;25(7):4096. DOI: 10.3390/ijms25074096.
40. Bonowicz K., Mikołajczyk K., Faisal I., Qamar M., Steinbrink K., Kleszczyński K. et al. Mechanism of extracellular vesicle secretion associated with TGF- β -dependent inflammatory response in the tumor microenvironment. *Int. J. Mol. Sci.* 2022;23(23):15335. DOI: 10.3390/ijms232315335.
41. Lin Y.C., Sahoo B.K., Gau S.S., Yang R.B. The biology of SCUBE. *J. Biomed Sci.* 2023;30(1):33. DOI: 10.1186/s12929-023-00925-3.
42. Lin Y.C., Chang Y.J., Gau S.S., Lo C.M., Yang R.B. et al. SCUBE2 regulates adherens junction dynamics and vascular barrier function during inflammation. *Cardiovasc. Res.* 2024;120(13):1636–1649. DOI: 10.1093/cvr/cvae132.
43. Benz K., Varga I., Neureiter D., Campean V., Daniel C., Heim C. et al. Vascular inflammation and media calcification are already present in early stages of chronic kidney disease. *Cardiovasc. Pathol.* 2017;27:57–67. DOI: 10.1016/j.carpath.2017.01.004.
44. Jansen I., Cahalane R., Hengst R., Akyildiz A., Farrell E., Gijzen F. et al. The interplay of collagen, macrophages, and microcalcification in atherosclerotic plaque cap rupture mechanics. *Basic Res. Cardiol.* 2024;119(2):193–213. DOI: 10.1007/s00395-024-01033-5.
45. Akers E.J., Nicholls S.J., Di Bartolo B.A. Plaque calcification: do lipoproteins have a role? *Arterioscler. Thromb. Vasc. Biol.* 2019;39(10):1902–1910. DOI: 10.1161/ATVBAHA.119.311574.
46. Shioi A., Ikari Y. Plaque calcification during atherosclerosis progression and regression. *J. Atheroscler. Thromb.* 2018;25(4):294–303. DOI: 10.5551/jat.RV17020.
47. Woo S.H., Kim D.Y., Choi J.H. Roles of vascular smooth muscle cells in atherosclerotic calcification. *J. Lipid. Atheroscler.* 2023;12(2):106–118. DOI: 10.12997/jla.2023.12.2.106.
48. Zhang F., Guo X., Xia Y., Mao L. An update on the phenotypic switching of vascular smooth muscle cells in the pathogenesis of atherosclerosis. *Cell Mol. Life Sci.* 2021;79(1):6. DOI: 10.1007/s00018-021-04079-z.
49. Speer M.Y., Li X., Hiremath P.G., Giachelli C.M. Runx2/Cbfa1, but not loss of myocardium, is required for smooth muscle cell lineage reprogramming toward osteochondrogenesis. *J. Cell Biochem.* 2010;110(4):935–947. DOI: 10.1002/jcb.22607.
50. Segura A.M., Radovancevic R., Connelly J.H., Loyalka P., Gregoric I.D., Buja L.M. et al. Endomyocardial nodular calcification as a cause of heart failure. *Cardiovasc. Pathol.* 2011;20(5):e185–e188. DOI: 10.1016/j.carpath.2010.08.003.
51. Zazzeroni L., Faggioli G., Pasquinelli G. Mechanisms of arterial calcification: the role of matrix vesicles. *Eur. J. Vasc. Endovasc. Surg.* 2018;55(3):425–432. DOI: 10.1016/j.ejvs.2017.12.009.
52. Polonskaya Ya.V., Kashtanova E.V., Stakhneva E.M., Ledovskikh S.R., Garbuzova E.V., Shramko V.S. et al. Levels of metalloproteinases and adipose tissue hormones in men with coronary atherosclerosis. *Bulletin of Siberian Medicine*. 2023;22(4):73–78. (In Russ.). DOI: 10.20538/1682-0363-2023-4-73-78.
53. Polonskaya Ya.V., Ragino Yu.I. Metalloproteinase and atherosclerosis. *Atherosclerosis*. 2017;13(3):50–55. (In Russ.).
54. Volkov A.M., Murashov I.S., Polonskaya Y.V., Savchenko S.V., Kazanskaya G.M., Kliver E.E., Chernyavsky A.M. Changes in Matrix Metalloproteinase Content and Their Tissue Expression in Atherosclerotic Plaques of Different Types. *Cardiology*. 2018;58(10):12–8. (In Russ.). DOI: 10.18087/cardio.2018.10.10180.
55. Polonskaya Y.V., Kashtanova E.V., Stakhneva E.M., Sadvovsky E.V., Ragino Y.I. The Role of Matrix Metalloproteinases and Tissue Inhibitors of Metalloproteinases in the Development of Coronary Atherosclerosis. *Atherosclerosis*. 2021;17(3):76–78. (In Russ.). DOI: 10.52727/2078-256X-2021-17-3-76-78.
56. Olejars W., Łacheta D., Kubiak-Tomaszewska G. Matrix metalloproteinases as biomarkers of atherosclerotic plaque instability. *Int. J. Mol. Sci.* 2020;21(11):3946. DOI: 10.3390/ijms21113946.
57. Samah N., Ugusman A., Hamid A.A., Sulaiman N., Aminuddin A. Role of matrix metalloproteinase-2 in the development of atherosclerosis among patients with coronary artery disease. *Mediators Inflamm.* 2023;2023:9715114. DOI: 10.1155/2023/9715114.
58. Yang C., Qiao S., Song Y., Liu Y., Tang Y., Deng L. et al. Procollagen type I carboxy-terminal propeptide (PICP) and MMP-2 are potential biomarkers of myocardial fibrosis in patients with hypertrophic cardiomyopathy. *Cardiovasc. Pathol.* 2019;43:107150. DOI: 10.1016/j.carpath.2019.107150.
59. Bräuninger H., Krüger S., Bacmeister L., Nyström A., Eyerich K., Westermann D. et al. Matrix metalloproteinases in coronary artery disease and myocardial infarction. *Basic Res. Cardiol.* 2023;118(1):18. DOI: 10.1007/s00395-023-00987-2.
60. Shakhshneider E.V., Ivanoshchuk D.E., Ragino Yu.I., Fishman V.S., Polonskaya Ya.V., Kashtanova E.V. et al. Analysis of differential expression of lipid metabolism genes in ath-

- erosclerotic plaques in patients with coronary atherosclerosis. *Siberian Journal of Clinical and Experimental Medicine*. 2021;36(4):156–163. DOI: 10.29001/2073-8552-2021-36-4-156-163.
61. Garbuzova E.V., Polonskaya Ya.V., Kashtanova E.V., Stakhneva E.M., Shramko V.S., Murashov I.S. et al. Biomolecules of adipose tissue in atherosclerotic plaques of men with coronary atherosclerosis. *Kardiologiya*. 2024;64(8):39–47. DOI: 10.18087/cardio.2024.8.n2634.
 62. Luo X., Lv Y., Bai X., Qi J., Weng X., Liu S. et al. Plaque erosion: A distinctive pathological mechanism of acute coronary syndrome. *Front. Cardiovasc. Med*. 2021;8:711453. DOI: 10.3389/fcvm.2021.711453.
 63. Fahed A.C., Jang I.K. Plaque erosion and acute coronary syndromes: phenotype, molecular characteristics and future directions. *Nat. Rev. Cardiol*. 2021;18:724–734. DOI: 10.1038/s41569-021-00542-3.
 64. Lv Y., Pang X., Cao Z., Song C., Liu B., Wu W. et al. Evolution and function of the notch signaling pathway: an invertebrate perspective. *Int. J. Mol. Sci*. 2024;25(6):3322. DOI: 10.3390/ijms25063322.
 65. Rizzo P., Ferrari R. The Notch pathway: a new therapeutic target in atherosclerosis? *Eur. Heart J. Suppl.* 2015;17 (Suppl. _A):A74–A76. DOI: 10.1093/eurheartj/suv011.
 66. Suarez Rodriguez F., Sanlidag S., Sahlgren C. Mechanical regulation of the notch signaling pathway. *Curr. Opin. Cell Biol*. 2023;85:102244. DOI: 10.1016/j.ccb.2023.102244.
 67. Vicceli Dalla Sega F., Fortini F., Aquila G., Campo G., Vaccarezza M., Rizzo P. Notch signaling regulates immune responses in atherosclerosis. *Front. Immunol*. 2019;10:1130. DOI: 10.3389/fimmu.2019.01130.
 68. Souilhol C., Tardajos Ayllon B., Li X. JAG1-NOTCH4 mechanosensing drives atherosclerosis. *Sci. Adv*. 2022;8(35):eabo7958. DOI: 10.1126/sciadv.abo7958.
 69. Harrison O.J., Visan A.C., Moorjani N. Defective NOTCH signaling drives increased vascular smooth muscle cell apoptosis and contractile differentiation in bicuspid aortic valve aortopathy: A review of the evidence and future directions. *Trends Cardiovasc. Med*. 2019;29(2):61–68. DOI: 10.1016/j.tcm.2018.06.008.
 70. Mao C., Ma Z., Jia Y., Li W., Xie N., Zhao G. et al. Nidogen-2 maintains the contractile phenotype of vascular smooth muscle cells and prevents neointima formation via bridging jagged1-notch3 signaling. *Circulation*. 2021;144(15):1244–1261. DOI: 10.1161/CIRCULATIONAHA.120.053361.
 71. Kocelak P., Puzianowska-Kuźnicka M., Olszanecka-Glinianowicz M., Chudek J. Wnt signaling pathway and sclerostin in the development of atherosclerosis and vascular calcification. *Adv. Clin. Exp. Med*. 2024;33(5):519–532. DOI: 10.17219/acem/169567.
 72. Abou Azar F., Lim G.E. Metabolic contributions of wnt signaling: more than controlling flight. *Front. Cell Dev. Biol*. 2021;9:709823. DOI: 10.3389/fcell.2021.709823.
 73. Wang K., Zhang R., Lehwald N., Tao G.Z., Liu B., Liu B. et al. Wnt/β-catenin signaling activation promotes lipogenesis in the steatotic liver via physical mTOR interaction. *Front. Endocrinol. (Lausanne)*. 2023;14:1289004. DOI: 10.3389/fendo.2023.1289004.
 74. Weinstock A., Rahman K., Yaacov O., Nishi H., Menon P., Nikain C.A. et al. Wnt signaling enhances macrophage responses to IL-4 and promotes resolution of atherosclerosis. *Elife*. 2021;10:e67932. DOI: 10.7554/eLife.67932.
 75. Zhang Y., Wu H., He R., Ye C., Chen H., Wang J. et al. Dickkopf-2 knockdown protects against classic macrophage polarization and lipid loading by activation of Wnt/β-catenin signaling. *J. Cardiol*. 2021;78(4):328–333. DOI: 10.1016/j.jjcc.2021.04.010.
 76. Hassanabad A.F., Zarzycki A.N., Patel V.B., Fedak P.W.M. Current concepts in the epigenetic regulation of cardiac fibrosis. *Cardiovasc. Pathol*. DOI: 10.1016/j.carpath.2024.107673.
 77. Wang J., Hu X., Hu X., Gao F., Li M., Cui Y. et al. MicroRNA-520c-3p targeting of RelA/p65 suppresses atherosclerotic plaque formation. *Int. J. Biochem. Cell Biol*. 2021;131:105873. DOI: 10.1016/j.biocel.2020.105873.
 78. Li Z., Xu C., Sun D. MicroRNA-488 serves as a diagnostic marker for atherosclerosis and regulates the biological behavior of vascular smooth muscle cells. *Bioengineered*. 2021;12(1):4092–4099. DOI: 10.1080/21655979.2021.1953212.
 79. Lv D., Guo Y., Zhang L., Li X., Li G. Circulating miR-183-5p levels are positively associated with the presence and severity of coronary artery disease. *Front. Cardiovasc. Med*. 2023;10:1196348. DOI: 10.3389/fcvm.2023.1196348.
 80. Kim M., Zhang X. The profiling and role of miRNAs in diabetes mellitus. *J. Diabetes Clin. Res*. 2019;1(1):5–23. DOI: 10.33696/diabetes.1.003.
 81. Jiang Q., Li Y., Wu Q., Huang L., Xu J., Zeng Q. Pathogenic role of microRNAs in atherosclerotic ischemic stroke: implications for diagnosis and therapy. *Genes Dis*. 2021;9(3):682–696. DOI: 10.1016/j.gendis.2021.01.001.
 82. Mahjoubin-Tehran M., Aghaee-Bakhtiari S.H., Sahebkar A., Butler A.E., Oskuee R.K. *In silico* and *in vitro* analysis of microRNAs with therapeutic potential in atherosclerosis. *Sci. Rep*. 2022;12(1):20334. DOI: 10.1038/s41598-022-24260-z.
 83. Peng Q., Yin R., Zhu X., Jin L., Wang J., Pan X. et al. miR-155 activates the NLRP3 inflammasome by regulating the MEK/ERK/NF-κB pathway in carotid atherosclerotic plaques in ApoE^{-/-} mice. *J. Physiol. Biochem*. 2022;78(2):365–375. DOI: 10.1007/s13105-022-00871-y.
 84. Vavlukis M., Vavlukis A. Adding ezetimibe to statin therapy: latest evidence and clinical implications. *Drugs Context*. 2018;7:212534. DOI: 10.7573/dic.212534.
 85. Tricarico P.M., Crovella S., Celsi F. Mevalonate pathway blockade, mitochondrial dysfunction and autophagy: a possible link. *Int. J. Mol. Sci*. 2015;16(7):16067–16084. DOI: 10.3390/ijms160716067.
 86. Kim S.W., Kang H.J., Jhon M., Kim J.W., Lee J.Y., Walker A.J. et al. Statins and inflammation: new therapeutic opportunities in psychiatry. *Front. Psychiatry*. 2019;10:103. DOI: 10.3389/fpsyt.2019.00103.
 87. Koushki K., Shahbaz S.K., Mashayekhi K., Sadeghi M., Zayeri Z.D., Taba M.Y. et al. Anti-inflammatory action of statins in cardiovascular disease: the role of inflammasome and toll-like receptor pathways. *Clin. Rev. Allergy Immunol*. 2021;60(2):175–199. DOI: 10.1007/s12016-020-08791-9.
 88. Zivkovic S., Maric G., Cvetinovic N., Lepojevic-Stefanovic D., Bozic Cvijan B. Anti-Inflammatory Effects of Lipid-Lower-

- ing Drugs and Supplements-A Narrative Review. *Nutrients*. 2023;15(6):1517. DOI: 10.3390/nu15061517.
89. Frostegård J., Zhang Y., Sun J., Yan K., Liu A. Oxidized low-density lipoprotein (OxLDL)-treated dendritic cells promote activation of T cells in human atherosclerotic plaque and blood, which is repressed by statins: microRNA let-7c is integral to the effect. *J. Am. Heart Assoc.* 2016;5(9):e003976. DOI: 10.1161/JAHA.116.003976.
 90. Fogacci F., Giovannini M., Tocci G., Imbalzano E., Borghi C., Cicero A.F.G. Effect of Coenzyme Q10 on Physical Performance in Older Adults with Statin-Associated Asthenia: A Double-Blind, Randomized, Placebo-Controlled Clinical Trial. *J. Clin. Med.* 2024;13(13):3741. DOI: 10.3390/jcm13133741.
 91. Kobalava Zh.D., Lazarev P.V., Villevalde S.V. Statin-Associated Diabetes Mellitus: Status of the Problem in 2018. *Russian Journal of Cardiology*. 2018;(9):89-99. (In Russ.). DOI: 10.15829/1560-4071-2018-9-89-99.
 92. Cicero A.F.G., Morbini M., Bove M., D'Addato S., Fogacci F., Rosticci M. et al. Additional therapy for cholesterol lowering in ezetimibe-treated, statin-intolerant patients in clinical practice: results from an internal audit of a university lipid clinic. *Curr. Med. Res. Opin.* 2016;32(10):1633–1638. DOI: 10.1080/03007995.2016.1190326.
 93. Strilchuk L., Tocci G., Fogacci F., Cicero A.F.G. An overview of rosuvastatin/ezetimibe association for the treatment of hypercholesterolemia and mixed dyslipidemia. *Expert Opin. Pharmacother.* 2020;21(5):531–539. DOI: 10.1080/14656566.2020.1714028.
 94. Fogacci F., Borghi C., Cicero A.F.G. Misinterpreting data in lipidology in the era of COVID-19. *J. Clin. Lipidol.* 2020;14(4):543–544. DOI: 10.1016/j.jacl.2020.07.004.
 95. Mohseni M., Mohammadifard N., Hassannejad R., Aghabozorgi M., Shirani F., Sadeghi M. et al. Longitudinal association of dietary habits and the risk of cardiovascular disease among Iranian population between 2001 and 2013: the Isfahan Cohort Study. *Sci. Rep.* 2023;13(1):5364. DOI: 10.1038/s41598-023-32387-w.
 96. Pauley M.E., Vinovskis C., MacDonald A., Baca M., Pyle L., Wadwa R.P. et al. Triglyceride content of lipoprotein subclasses and kidney hemodynamic function and injury in adolescents with type 1 diabetes. *J. Diabetes Complications*. 2023;37(2):108384. DOI: 10.1016/j.jdiacomp.2022.108384.
 97. Liu H.H., Li S., Cao Y.X., Guo Y.L., Zhu C.G., Wu N.Q. et al. Association of triglyceride-rich lipoprotein-cholesterol with recurrent cardiovascular events in statin-treated patients according to different inflammatory status. *Atherosclerosis*. 2021;330:29–35. DOI: 10.1016/j.atherosclerosis.2021.06.907.
 98. Fogacci F., Yerlitaş S.İ., Giovannini M., Zararsız G., Lido P., Borghi C. et al. Sex X time interactions in Lp(a) and LDL-C response to evolocumab. *Biomedicines*. 2023;11(12):3271. DOI: 10.3390/biomedicines11123271.
 99. Cicero A.F.G., Toth P.P., Fogacci F., Virdis A., Borghi C. Improvement in arterial stiffness after short-term treatment with PCSK9 inhibitors. *Nutr. Metab. Cardiovasc. Dis.* 2019;29(5):527–529. DOI: 10.1016/j.numecd.2019.01.010.
 100. Cicero A.F.G., Fogacci F., Bragagni A., Borghi C. Short-term evolocumab-induced tendon xanthomas regression in an elderly patient with homozygous familial hypercholesterolemia. *Intern. Emerg. Med.* 2023;18(1):307–310. DOI: 10.1007/s11739-022-03106-6.
 101. Sabatine M.S., Giugliano R.P., Keech A.C., Honarpour N., Wiviott S.D., Murphy S.A. et al. Evolocumab and clinical outcomes in patients with cardiovascular disease. *N. Engl. J. Med.* 2017;376(18):1713–1722. DOI: 10.1056/NEJMoa1615664.
 102. Schwartz G.G., Steg P.G., Szarek M., Bhatt D.L., Bittner V.A., Diaz R. et al. Alirocumab and cardiovascular outcomes after acute coronary syndrome. *N. Engl. J. Med.* 2018;379(22):2097–2107. DOI: 10.1056/NEJMoa1801174.
 103. Bodapati A.P., Hanif A., Okafor D.K., Katyal G., Kaur G., Ashraf H. et al. PCSK-9 Inhibitors and Cardiovascular Outcomes: A Systematic Review With Meta-Analysis. *Cureus*. 2023;15(10):e46605. DOI: 10.7759/cureus.46605.
 104. Khan S.U., Talluri S., Riaz H., Rahman H., Nasir F., Bin Riaz I. et al. A Bayesian network meta-analysis of PCSK9 inhibitors, statins and ezetimibe with or without statins for cardiovascular outcomes. *Eur. J. Prev. Cardiol.* 2018;25(8):844–853. DOI: 10.1177/2047487318766612.
 105. Namitokov A.M., Zafiraki V.K., Karabakhtsieva K.V. Perspectives of Using PCSK-9 Modifying Agents in Acute Coronary Syndrome. *Innovative Medicine of Kuban*. 2024;(2):124-128. (In Russ.). DOI: 10.35401/2541-9897-2024-9-2-124-128.
 106. Cheng J.M., Oemrawsingh R.M., Garcia-Garcia H.M. PCSK9 in relation to coronary plaque inflammation: Results of the ATHEROREMO-IVUS study. *Atherosclerosis*. 2016;248:117–122. DOI: 10.1016/j.atherosclerosis.2016.03.010.
 107. Almontashiri N.A., Vilmundarson R.O., Ghasemzadeh N., Dandona S., Roberts R., Quyyumi A.A. et al. Plasma PCSK9 levels are elevated with acute myocardial infarction in two independent retrospective angiographic studies. *PLoS One*. 2014;9(9):e106294. DOI: 10.1371/journal.pone.0106294.
 108. Badimon L., Luquero A., Crespo J., Peña E., Borrell-Pages M. PCSK9 and LRP5 in macrophage lipid internalization and inflammation. *Cardiovasc. Res.* 2021;117(9):2054–2068. DOI: 10.1093/cvr/cvaa254.
 109. Wu N.Q., Shi H.W., Li J.J. Proprotein Convertase Subtilisin/Kexin Type 9 and Inflammation: An Updated Review. *Front. Cardiovasc. Med.* 2022;9:763516. DOI: 10.3389/fcvm.2022.763516.
 110. Fruchart Gaillard C., Ouadda A.B.D., Ciccone L., Girard E., Mikaeeli S., Evagelidis A. et al. Molecular interactions of PCSK9 with an inhibitory nanobody, CAP1 and HLA-C: functional regulation of LDLR levels. *Mol. Metab.* 2023;67:101662. DOI: 10.1016/j.molmet.2022.101662.
 111. Shin D., Kim S., Lee H., Lee J., Park H.W. PCSK9 stimulates Syk, PKC δ , and NF- κ B, leading to atherosclerosis progression independently of LDL receptor. *Nat. Commun.* 2024;15(1):2789. DOI: 10.1038/s41467-024-46336-2.
 112. Banerjee Y., Pantea Stoian A., Cicero A.F.G., Fogacci F., Nikolic D., Sachinidis A. et al. Inclisiran: a small interfering RNA strategy targeting PCSK9 to treat hypercholesterolemia. *Expert Opin. Drug Saf.* 2022;21(1):9–20. DOI: 10.1080/14740338.2022.1988568.
 113. Cicero A.F.G., Fogacci F., Zambon A., Toth P.P., Borghi C.

- Efficacy and safety of inclisiran a newly approved FDA drug: a systematic review and pooled analysis of available clinical studies. *Am. Heart J. Plus.* 2022;13:100127. DOI: 10.1016/j.ahjo.2022.100127.
114. Strilchuk L., Fogacci F., Cicero A.F. Safety and tolerability of injectable lipid-lowering drugs: an update of clinical data. *Expert Opin. Drug Saf.* 2019;18(7):611–621. DOI: 10.1080/14740338.2019.1620730.
 115. Voevoda M.I., Gurevich V.S., Ezhov M.V., Sergienko I.V. Inclisiran – A New Era in Hypolipidemic Therapy. *Cardiology.* 2022;62(6):57–62. (In Russ.). DOI: 10.18087/cardio.2022.6.n2115.
 116. Khan S.A., Naz A., Qamar Masood M., Shah R. Meta-Analysis of Inclisiran for the Treatment of Hypercholesterolemia. *Am. J. Cardiol.* 2020;134:69–73. DOI: 10.1016/j.amjcard.2020.08.018.
 117. Fogacci F., Di Micoli V., Sabouret P., Giovannini M., Cicero A.F.G. Lifestyle and lipoprotein(a) levels: does a specific counseling make sense? *J. Clin. Med.* 2024;13(3):751. DOI: 10.3390/jcm13030751.
 118. Fogacci F., Di Micoli V., Avagimyan A., Giovannini M., Imbalzano E., Cicero A.F.G. Assessment of apolipoprotein(a) isoform size using phenotypic and genotypic methods. *Int. J. Mol. Sci.* 2023;24(18):13886. DOI: 10.3390/ijms241813886.
 119. Dzobo K.E., Kraaijenhof J.M., Stroes E.S.G., Nurmohamed N.S., Kroon J. Lipoprotein(a): an underestimated inflammatory mastermind. *Atherosclerosis.* 2022;349:101–109. DOI: 10.1016/j.atherosclerosis.2022.04.004.
 120. Gianos E., Duell P.B., Toth P.P., Moriarty P.M., Thompson G.R., Brinton E.A. et al. American Heart Association Council on Arteriosclerosis, Thrombosis and Vascular Biology; Council on Lifelong Congenital Heart Disease and Heart Health in the Young; and Council on Peripheral Vascular Disease. Lipoprotein Apheresis: Utility, Outcomes, and Implementation in Clinical Practice: A Scientific Statement From the American Heart Association. *Arterioscler. Thromb. Vasc. Biol.* 2024. DOI: 10.1161/ATV.0000000000000177.
 121. Di Fusco S.A., Maggioni A.P., Scicchitano P., Zuin M., D’Elia E., Colivicchi F. Lipoprotein (a), inflammation, and atherosclerosis. *J. Clin. Med.* 2023;12(7):2529. DOI: 10.3390/jcm12072529.
 122. Lampsas S., Xenou M., Oikonomou E., Pantelidis P., Lysandrou A., Sarantos S. et al. Lipoprotein(a) in atherosclerotic diseases: from pathophysiology to diagnosis and treatment. *Molecules.* 2023;28(3):969. DOI: 10.3390/molecules28030969.
 123. Afzal Z., Cao H., Chaudhary M., Chigurupati H.D., Nepala S., Alruwaili W. et al. Elevated lipoprotein(a) levels: A crucial determinant of cardiovascular disease risk and target for emerging therapies. *Curr. Probl. Cardiol.* 2024;49(8):102586. DOI: 10.1016/j.cpcardiol.2024.102586.
 124. Schnitzler J.G., Hoogeveen R.M., Ali L., Prange K.H.M., Waissi F., van Weeghel M. et al. Atherogenic lipoprotein(a) increases vascular glycolysis, thereby facilitating inflammation and leukocyte extravasation. *Circ. Res.* 2020;126(10):1346–1359. DOI: 10.1161/CIRCRESA-HA.119.316206.
 125. Karwatowska-Prokopczuk E., Lesogor A., Yan J.H., Hurh E., Hoenlinger A., Margolske A. et al. Efficacy and safety of pelacarsen in lowering Lp(a) in healthy Japanese subjects. *J. Clin. Lipidol.* 2023;17(1):181–188. DOI: 10.1016/j.jacl.2022.12.001.
 126. O’Donoghue M.L., Rosenson R.S., Gencer B., López J.A.G., Lepor N.E., Baum S.J. et al. Small interfering RNA to reduce lipoprotein(a) in cardiovascular disease. *N. Engl. J. Med.* 2022;387(20):1855–1864. DOI: 10.1056/NEJMoa2211023.
 127. Nissen S.E., Wolski K., Watts G.F., Koren M.J., Fok H., Nicholls S.J., Rider D.A. et al. Single Ascending and Multiple-Dose Trial of Zelasiran, a Short Interfering RNA Targeting Lipoprotein(a): A Randomized Clinical Trial. *JAMA.* 2024;331(18):1534–1543. DOI: 10.1001/jama.2024.4504.
 128. Nissen S.E., Linnebjerg H., Shen X., Wolski K., Ma X., Lim S. et al. Lepodisiran, an Extended-Duration Short Interfering RNA Targeting Lipoprotein(a): A Randomized Dose-Ascending Clinical Trial. *JAMA.* 2023;330(21):2075–2083. DOI: 10.1001/jama.2023.21835.
 129. Nicholls S.J., Nissen S.E., Fleming C., Urva S., Suico J., Berg P.H. et al. Muvalaplin, an Oral Small Molecule Inhibitor of Lipoprotein(a) Formation: A Randomized Clinical Trial. *JAMA.* 2023;330(11):1042–1053. DOI: 10.1001/jama.2023.16503.
 130. O’Donoghue M.L., Rosenson R.S., López J.A.G., Lepor N.E., Baum S.J., Stout E. et al. The off-treatment effects of olpasiran on lipoprotein(a) lowering: OCEAN(a)-dose extension period results. *J. Am. Coll. Cardiol.* 2024;84(9):790–797. DOI: 10.1016/j.jacc.2024.05.058.
 131. Nicholls S.J., Ni W., Rhodes G.M., Nissen S.E., Navar A.M., Michael L.F. et al. Oral Muvalaplin for Lowering of Lipoprotein(a): A Randomized Clinical Trial. *JAMA.* 2024:e2424017. DOI: 10.1001/jama.2024.24017.
 132. Nissen S.E., Wang Q., Nicholls S.J., Navar A.M., Ray K.K., Schwartz G.G. et al. Zelasiran-A Small-Interfering RNA Targeting Lipoprotein(a): A Phase 2 Randomized Clinical Trial. *JAMA.* 2024:e2421957. DOI: 10.1001/jama.2024.21957.
 133. Kolovou G., Kolovou V., Katsiki N. Volanesorsen: A new era in the treatment of severe hypertriglyceridemia. *J. Clin. Med.* 2022;11(4):982. DOI: 10.3390/jcm11040982.
 134. Stroes E.S.G., Alexander V.J., Karwatowska-Prokopczuk E. Olezarsen, acute pancreatitis, and familial chylomicronemia syndrome. *N. Engl. J. Med.* 2024;390(19):1781–1792. DOI: 10.1056/NEJMoa2400201.
 135. Witztum J.L., Gaudet D., Freedman S.D., Alexander V.J., Digenio A., Williams K.R. et al. Volanesorsen and triglyceride levels in familial chylomicronemia syndrome. *N. Engl. J. Med.* 2019;381(6):531–542. DOI: 10.1056/NEJMoa1715944.
 136. Witztum J.L., Gaudet D., Arca M., Jones A., Soran H., Gouni-Berthold I. et al. Corrigendum to Volanesorsen and triglyceride levels in familial chylomicronemia syndrome: Long-term efficacy and safety data from patients in an open-label extension trial. *J. Clin. Lipidol.* 2024;18(3):e482–e483. DOI: 10.1016/j.jacl.2023.09.010.
 137. Gouni-Berthold I., Alexander V.J., Yang Q., Hurh E., Steinhagen-Thiessen E., Moriarty P.M. et al. Efficacy and safety of volanesorsen in patients with multifactorial chylomicronaemia (COMPASS): a multicentre, double-blind, randomised, placebo-controlled, phase 3 trial. *Lancet Diabetes Endocrinol.*

- 2021;9(5):264–275. DOI: 10.1016/S2213-8587(21)00046-2.
138. Hegele R.A. Apolipoprotein C-III inhibition to lower triglycerides: one ring to rule them all? *Eur. Heart J.* 2022;43(14):1413–1415. DOI: 10.1093/eurheartj/ehab890.
 139. Tardif J.C., Karwowska-Prokopczuk E., Amour E.S., Ballantyne C.M., Shapiro M.D., Moriarty P.M. et al. Apolipoprotein C-III reduction in subjects with moderate hypertriglyceridaemia and at high cardiovascular risk. *Eur. Heart J.* 2022;43(14):1401–1412. DOI: 10.1093/eurheartj/ehab820.
 140. Petrosyan A.S., Rudy R.S., Polyakov P.P., Kade A.H., Zanin S.A. Pathogenetic Mechanisms of the Effects of Bempedoic Acid. *Rational Pharmacotherapy in Cardiology.* 2022;18(6):734–741. (In Russ.). DOI: 10.20996/1819-6446-2022-12-11
 141. Ruscica M., Sirtori C.R., Carugo S., Banach M., Corsini A. Bempedoic Acid: for Whom and When. *Curr Atheroscler Rep.* 2022;24(10):791–801. DOI: 10.1007/s11883-022-01054-2.
 142. Mazerkina I.A., Bukatina T.M., Aleksandrova T.V. Efficacy and Safety of Bempedoic Acid as a New Hypolipidemic Agent. *Safety and Risk of Pharmacotherapy.* 2023;11(3):292–302. (In Russ.). DOI: 10.30895/2312-7821-2023-11-3-292-302.
 143. Ballantyne C.M., Banach M., Mancini G.B.J., Lepor N.E., Hanselman J.C., Zhao X. et al. Efficacy and safety of bempedoic acid added to ezetimibe in statin-intolerant patients with hypercholesterolemia: A randomized, placebo-controlled study. *Atherosclerosis.* 2018;277:195–203. DOI: 10.1016/j.atherosclerosis.2018.06.002.
 144. Ray K.K., Bays H.E., Catapano A.L., Lalwani N.D., Bloedon L.T., Sterling L.R. et al. Safety and efficacy of bempedoic acid to reduce LDL cholesterol. *N. Engl. J. Med.* 2019;380(11):1022–1032. DOI: 10.1056/NEJMoa1803917.
 145. Goldberg A.C., Leiter L.A., Stroses E.S.G., Baum S.J., Hanselman J.C., Bloedon L.T. et al. Effect of Bempedoic Acid vs Placebo Added to Maximally Tolerated Statins on Low-Density Lipoprotein Cholesterol in Patients at High Risk for Cardiovascular Disease: The CLEAR Wisdom Randomized Clinical Trial. *JAMA.* 2019;322(18):1780–1788. DOI: 10.1001/jama.2019.16585.
 146. Laufs U., Banach M., Mancini G.B.J., Gaudet D., Bloedon L.T., Sterling L.R. et al. Efficacy and safety of bempedoic acid in patients with hypercholesterolemia and statin intolerance. *J. Am. Heart Assoc.* 2019;8(7):e011662. DOI: 10.1161/JAHA.118.011662.
 147. Pinkosky S.L., Filippov S., Srivastava R.A., Hanselman J.C., Bradshaw C.D., Hurley T.R. et al. AMP-activated protein kinase and ATP-citrate lyase are two distinct molecular targets for ETC-1002, a novel small molecule regulator of lipid and carbohydrate metabolism. *J. Lipid Res.* 2013;54(1):134–151. DOI: 10.1194/jlr.M030528.
 148. Pinkosky S.L., Newton R.S., Day E.A., Ford R.J., Lhotak S., Austin R.C., Birch C.M. et al. Liver-specific ATP-citrate lyase inhibition by bempedoic acid decreases LDL-C and attenuates atherosclerosis. *Nat. Commun.* 2016;7:13457. DOI: 10.1038/ncomms13457.
 149. Morrow M.R., Batchuluun B., Wu J., Ahmadi E., Leroux J.M., Mohammadi-Shemirani P. et al. Inhibition of ATP-citrate lyase improves NASH, liver fibrosis, and dyslipidemia. *Cell Metab.* 2022;34(6):919–936.e8. DOI: 10.1016/j.cmet.2022.05.004.
 150. Raal F.J., Rosenson R.S., Reeskamp L.F., Hovingh G.K., Kastelein J.J.P., Rubba P. et al. Evinacumab for homozygous familial hypercholesterolemia. *N. Engl. J. Med.* 2020;383(8):711–720. DOI: 10.1056/NEJMoa2004215.
 151. Gill P.K., Hegele R.A. New biological therapies for low-density lipoprotein cholesterol. *Can. J. Cardiol.* 2023;39(12):1913–1930. DOI: 10.1016/j.cjca.2023.08.003.
 152. Esba L.C.A., Alharbi H. Lomitapide: a medication use evaluation and a formulary perspective. *Glob. J. Qual. Saf. Healthc.* 2024;7(2):59–62. DOI: 10.36401/JQSH-23-32.
 153. Dybiec J., Baran W., Dąbek B., Fularski P., Młynarska E., Radzioch E. et al. Advances in treatment of dyslipidemia. *Int. J. Mol. Sci.* 2023;24(17):13288. DOI: 10.3390/ijms241713288.
 154. Sopert J., Lehrke M., Marx N., Jankowski J., Noels H. Lipoproteins and lipids in cardiovascular disease: from mechanistic insights to therapeutic targeting. *Adv. Drug Deliv. Rev.* 2020;159:4–33. DOI: 10.1016/j.addr.2020.07.019.
 155. Bagheri Kholenjani F., Shahidi S., Vaseghi G., Ashoorion V., Sarrafzadegan N., Siavash M. et al. First Iranian guidelines for the diagnosis, management, and treatment of hyperlipidemia in adults. *J. Res. Med. Sci.* 2024;29:18. DOI: 10.4103/jrms.jrms_318_23.
 156. Leung Y.Y., Yao Hui L.L., Kraus V.B. Colchicine--Update on mechanisms of action and therapeutic uses. *Semin. Arthritis Rheum.* 2015;45(3):341–350. DOI: 10.1016/j.semarthrit.2015.06.013.
 157. Zhou H., Khan D., Gerdes N., Hagenbeck C., Rana M., Cornelius J.F. et al. Colchicine protects against ethanol-induced senescence and senescence-associated secretory phenotype in endothelial cells. *Antioxidants (Basel).* 2023;12(4):960. DOI: 10.3390/antiox12040960.
 158. Aldana-Bitar J., Golub I.S., Moore J., Krishnan S., Verghese D., Manubolu V.S. et al. Colchicine and plaque: a focus on atherosclerosis imaging. *Prog. Cardiovasc. Dis.* 2024;84:68–75. DOI: 10.1016/j.pcad.2024.02.010.
 159. Tucker B., Goonetilleke N., Patel S., Keech A. Colchicine in atherosclerotic cardiovascular disease. *Heart.* 2024;110(9):618–625. DOI: 10.1136/heartjnl-2023-323177.
 160. Zhang F.S., He Q.Z., Qin C.H., Little P.J., Weng J.P., Xu S.W. Therapeutic potential of colchicine in cardiovascular medicine: a pharmacological review. *Acta Pharmacol. Sin.* 2022;43(9):2173–2190. DOI: 10.1038/s41401-021-00835-w.
 161. Tardif J.C., Kouz S., Waters D.D., Bertrand O.F., Diaz R., Maggioni A.P. et al. Efficacy and safety of low-dose colchicine after myocardial infarction. *N. Engl. J. Med.* 2019;381(26):2497–2505. DOI: 10.1056/NEJMoa1912388.
 162. Nidorf S.M., Fiolet A.T.L., Mosterd A., Eikelboom J.W., Schut A., Opstal T.S.J. et al. Colchicine in patients with chronic coronary disease. *N. Engl. J. Med.* 2020;383(19):1838–1847. DOI: 10.1056/NEJMoa2021372.
 163. Bresson D., Roubille F., Prieur C., Biere L., Ivanov F., Bouleti C. et al. Colchicine for Left Ventricular Infarct Size Reduction in Acute Myocardial Infarction: A Phase II, Multi-center, Randomized, Double-Blinded, Placebo-Controlled Study Protocol – The COVERT-MI Study. *Cardiology.* 2021;146(2):151–160. DOI: 10.1159/000512772.

164. Kelly P., Lemmens R., Weimar C., Walsh C., Purroy F., Barber M. et al. Long-term colchicine for the prevention of vascular recurrent events in non-cardioembolic stroke (CONVINCE): a randomised controlled trial. *Lancet*. 2024;404(10448):125–133. DOI: 10.1016/S0140-6736(24)00968-1.
165. Fogacci F., Al Ghasab N.S., Di Micoli V., Giovannini M., Cicero A.F.G. Cholesterol-lowering bioactive foods and nutraceuticals in pediatrics: clinical evidence of efficacy and safety. *Nutrients*. 2024;16(10):1526. DOI: 10.3390/nu16101526.
166. Cicero A.F.G., Fogacci F. The year in nutrition medicine 2023. *Arch. Med. Sci.* 2023;19(6):1599–1601. DOI: 10.5114/aoms/174787.
167. Sesso H.D., Manson J.E., Aragaki A.K., Rist P.M., Johnson L.G., Friedenberg G. et al. Effect of cocoa flavanol supplementation for the prevention of cardiovascular disease events: the Cocoa Supplement and Multivitamin Outcomes Study (COSMOS) randomized clinical trial. *Am. J. Clin. Nutr.* 2022;115(6):1490–1500. DOI: 10.1093/ajcn/nqac055.
168. Osadnik T., Goławski M., Lewandowski P., Morze J., Osadnik K., Pawlas N. et al. A network meta-analysis on the comparative effect of nutraceuticals on lipid profile in adults. *Pharmacol. Res.* 2022;183:106402. DOI: 10.1016/j.phrs.2022.106402.

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