REVIEWS AND LECTURES



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The Individual Contribution of Fatty Acids to the Development of Cardiovascular Diseases

Shramko V.S., Kashtanova E.V., Stakhneva E.M., Polonskaya Yu.V., Ragino Yu. I.

Research Institute of Internal and Preventive Medicine, Branch of the Institute of Cytology and Genetics, Siberian Branch of Russian Academy of Sciences

175/1 Boris Bogatkov St., 630089 Novosibirsk, Russian Federation

ABSTRACT

Impaired fatty acid (FA) metabolism may be an important factor that increases the development and progression of atherosclerosis and related cardiovascular diseases (CVD). However, most of the research focuses on studying the influence of classification groups of FA. Therefore, the aim of this lecture was to present both pro- and anti-atherogenic functions of each FA. This paper considers up-to-date information about the effects of saturated (myristic (C 14:0), palmitic (C 16:0), stearic (C 18:0)), monounsaturated (palmitoleic (C 16:1), oleic (C 18:1)), and polyunsaturated (linoleic (C 18:2 omega-6), alpha-linolenic (C 18:3, omega-3), dihomo-gamma-linolenic (C 20:3, omega-6), arachidonic (C 20:4, omega-6), eicosapentaenoic (C 20:5 omega-3)) FAs on CVD. The accumulated data expand the understanding of the role of FAs in metabolic processes, which will allow us to move from fundamental research to practical aspects of the use of these substances in the treatment of CVD. In the future, these results can be used in the interpretation and prediction of changes in lipid metabolism disorders in CVD.

Keywords: fatty acids, lipids, cardiovascular diseases, blood, risk factors

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Вклад жирных кислот в развитие сердечно-сосудистых заболеваний

Шрамко В.С., Каштанова Е.В., Стахнёва Е.М., Полонская Я.В., Рагино Ю.И.

Научно-исследовательский институт терапии и профилактической медицины – филиал Федерального исследовательского центра «Институт цитологии и генетики СО РАН» (НИИТПМ – филиал ИЦиГ СО РАН)

Россия, 630089, г. Новосибирск, ул. Б. Богаткова, 175/1

РЕЗЮМЕ

Нарушение обмена жирных кислот (ЖК) может являться значимым фактором, потенциирующим развитие и прогрессирование атеросклероза и связанных с ним сердечно-сосудистых заболеваний (ССЗ). Тем не менее большинство исследований сосредоточены на изучении влияния классификационных групп ЖК. Поэтому цель настоящей лекции – представить как про-, так и антиатерогенные функции каждой жирной кислоты.

В настоящей работе рассмотрены современные сведения о влиянии насыщенных (миристиновой (С 14:0),

пальмитиновой (С 16:0), стеариновой (С 18:0)), мононенасыщенных (пальмитолеиновой (С 16:1), олеиновой (С 18:1)) и полиненасыщенных (линолевой (С 18:2, омега-6), альфа-линоленовой (С 18:3, омега-3), дигомо-гамма-линоленовой (С 20:3, омега-6), арахидоновой (С 20:4, омега-6), эйкозапентаеновой (С 20:5, омега-3), докозагексаеновой (С 22:6, омега-3)) жирных кислот на ССЗ. Накопленные данные расширяют представления о роли ЖК в метаболических процессах, что позволит перейти от фундаментальнопоисковых работ к практическим аспектам применения данных веществ в лечении ССЗ. В перспективе эти результаты могут быть использованы при интерпретации и прогнозировании изменений метаболических нарушений липидов при ССЗ.

Ключевые слова: жирные кислоты, липиды, сердечно-сосудистые заболевания, кровь, факторы риска

Конфликт интересов. Авторы декларируют отсутствие явных и потенциальных конфликтов интересов, связанных с публикацией настоящей статьи

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INTRODUCTION

The growing prevalence of chronic noncommunicable diseases, primarily cardiovascular diseases (CVDs), is a huge problem for the health care system [1]. Coronary heart disease (CHD) caused by atherosclerotic lesions of the coronary arteries is the main and most common nosology among CVDs [2]. For a long time, atherosclerosis can be asymptomatic, which is associated with a latent stage of the disease, in which morphological changes in the coronary arteries are already present [3]. However, following the growth of the atherosclerotic plaque, gradual stenosis of the coronary and other arteries occurs, leading to complications, such as myocardial infarction (MI), stroke, angina pectoris, cerebrovascular insufficiency, sudden cardiac death, etc. [4, 5]. At the same time, the rate of atherosclerosis progression is strictly individual, which necessitates preventive measures at the population and individual levels aimed at eliminating or minimizing the incidence of CVD and the associated loss of working capacity.

A growing body of evidence suggests that fatty acids (FAs) and their metabolites play an important role in atherogenesis [6]. In addition to their structural and/or energy functions, FAs are associated with the regulation of hemodynamics, inflammation, endothelial dysfunction, antioxidant defense, and other important biological processes [7, 8]. This is due to their chemical structure, showing differences for both saturated (SFA) and unsaturated FA (UNFA) [6]. Therefore, the aim of this lecture was to study the role of each FA on the risk of developing CVD.

It should be noted that FAs are divided into short-chain, medium-chain, and long-chain FAs based on the number of carbon atoms in their hydrocarbon chain. In addition, according to the presence and number of double bonds in their carbon chain, they can be classified into SFAs (contain no double bonds); monounsaturated FAs (MUFA) (contain one double bond), and polyunsaturated FAs (PUFA), whose structure contains two or more double bonds [9, 10].

SATURATED FATTY ACIDS

As important energy sources, long-chain SFAs can be incorporated into lipoproteins, circulate in the blood, be stored in fat depots, and be used to synthesize other lipid compounds in the body [11]. Currently, the relationship between tissue SFA levels and the risk of atherosclerotic CVD is widely studied, mainly because SFAs can increase low-density lipoprotein cholesterol (LDL-C) concentrations [12]. Nevertheless, there is growing evidence that individual SFAs generally have different biological functions [13].

The most common SFA in the human body is palmitic acid (C16:0), which is an important component of membrane, secretory, and transport lipids, so both deficiency and excess of this SFA are harmful [14–16]. It can enter the body with food or be formed by endogenous synthesis (i.e., *de novo* lipogenesis) as a result of excess energy intake from carbohydrates and/or proteins [17]. To date, the relationship between high levels of palmitic acid in the blood and the risk of developing CVD is beyond doubt. The clinical and observational data indicate that C16:0 may be

associated with adverse cardiovascular events, as well as with overall mortality [18–20]. A population-based study by C.L. Chei et al., which was an additional study to the CIRCS (Circulatory Risk in Communities Study, Japan) [21], revealed that the average level of palmitic SFA was higher in patients with CAD than in the control group. Another population-based study, the LURIC (The Ludwigshafen Risk and Cardiovascular Health study, Germany) [19], showed a direct association with an increased risk of CVD mortality only for C16:0. Moreover, high palmitic acid intake (\approx 50% of total SFA intake) has been shown to elevate LDL-C [22] and interleukin-6 [18] levels and increase the risk of CHD [23, 24].

Stearic acid (C18:0) is also one of the main SFAs included in triglycerides. It can be obtained from a wide range of foods, including meat, fish, dairy products, etc. Meanwhile, under the action of palmitoyl elongase, cells can elongate C16:0 palmitic SFA to C18:0 stearic SFA [17]. Unlike palmitic FA, data on the effect of stearic SFA on lipid metabolism and, therefore, on the risk of CVD remain controversial. In the Mendelian Randomization Study [25], it was shown that a genetic predisposition to higher levels of stearic SFA in plasma was positively associated with CVD, such as stroke and venous thromboembolism. The EPIC-Norfolk study (European Prospective Investigation into Cancer, UK) [26] found that the concentration of stearic SFA in plasma was positively associated with an increased risk of CHD. At the same time, the CHS study (Cardiovascular Health Study, USA) [27] reported an inverse relationship between high C18:0 levels and all-cause mortality among elderly individuals (over 65 years of age). When studying the effect of stearic SFA, it was found that intake of C18:0 could reduce the level of total cholesterol, LDL-C, high-density lipoprotein cholesterol (HDL-C), and apolipoprotein A1 in the blood serum, compared to palmitic SFA [28].

However, no significant effect on LDL-C and HDL-C levels has been found in previous studies [22]. The Nurses' Health Study, which included data from the Health Professionals Follow-up Study [24], showed that higher intake of stearic SFA was associated with an increased risk of developing CHD over 24–28 years of follow-up. On the contrary, in the EPIC-NL study (European Prospective Investigation into Cancer and Nutrition–Netherlands Cohort, Netherlands) [29], no significant contribution of stearic acid to the development and course of CHD was found. Thus, the effect of stearic SFA on lipid

metabolism, inflammation, and/or endothelial function is not uniform, and additional research in this area is certainly needed.

One of the less common SFAs is myristic acid (C14:0). At relatively low concentrations in the human body, it is also an important component of cell membranes and can systematically influence lipoprotein metabolism [30]. The amount of endogenously biosynthesized myristic FA from lauric acid (C12:0) following elongation or from palmitic SFA following peroxisomal β-oxidation is much smaller than the amount supplied by dietary sources [31]. Within the Ventimiglia di Sicilia Heart Study [32], it was found that the levels of myristic SFA in plasma were inversely correlated with HDL-C levels. The Verona Heart Study reported a strong positive relationship between myristic acid and plasma apolipoprotein CIII concentrations [30]. The study by S.O. Ebbesson et al. [33] showed positive associations between high plasma C14:0 levels and CVD risk factors: increased levels of triglycerides, LDL-C, blood pressure (BP), body mass index (BMI), plasma glucose, as well as an inverse relationship with HDL-C. In an additional study to CIRCS [21], it was noted that high serum levels of myristic SFA were associated with an increased risk of CHD.

Nevertheless, a few data suggest that morbidity and mortality from CVD depend not so much on the total amount of SFA consumed, but on their ratio to UNFA [34].

MONOUNSATURATED FATTY ACIDS

The interest in the role of MUFA is steadily growing. In addition to exogenous intake, MUFAs can be endogenously synthesized in the liver and adipose tissue using microsomal stearoyl-CoA desaturase-1 from precursors – SFA [35]. MUFAs can promote a healthy blood lipid profile, improve blood pressure, glycemic control, etc. [36]. However, the effect of MUFAs on inflammation has not been sufficiently studied. However, there is increasing evidence indicating a close relationship between MUFAs and anti-inflammatory conditions [37]. Some of the key MUFAs, from the standpoint of their functional role in the body, are considered to be omega-7 palmitoleic (C16:1) and omega-9 oleic (C18:1) acids.

Recently, palmitoleic MUFA has been considered as a lipid hormone (or lipokine) derived from adipocytes, which allows adipose tissue to regulate systemic metabolism, indicating its physiological significance [38]. It has been established that C16:1

can be detected as a cis- or trans-isomer and is also associated with cholesterol metabolism, insulin sensitivity, and hemostasis [39–41]. At the same time, its effect on the body, in particular on the cardiovascular system, is still controversial among researchers. The EPIC-Norfolk Study [26], which involved 25,639 people, found no relationship between the content of palmitoleic MUFA in plasma and CHD.

In another prospective study – CIRCS [21], involving 12,840 individuals, positive associations of serum palmitoleic MUFA levels with a higher risk of developing CHD were registered in both men and women. In a population-based study of 1,828 patients with MI and 1,828 controls [42], it was found that C16:1 in adipose tissue had an inverse relationship with acute MI. Most likely, the opposite conclusions are due to different patient samples and/ or the biomaterial used. At the same time, a significant number of researchers are inclined to believe that palmitoleic MUFA can have an anti-inflammatory effect [43] and even reduce harmful effects of SFA. In particular, C16:1 promotes differentiation of primary macrophages into the anti-inflammatory M2 phenotype, protecting against the pro-inflammatory effects of palmitic acid [44]. In addition, C16:1 can reduce the levels of pro-inflammatory cytokines produced lipopolysaccharide-stimulated by macrophages (interleukin-6/-8, tumor necrosis factor α) [45].

Oleic acid accounts for approximately 80% of MUFAs in plasma phospholipids. In the PREDIMED study (PREvención con DIeta MEDiterránea, Spain) [46], researchers wanted to demonstrate that consumption of a Mediterranean diet enriched with olive oil (as a key component and source of plant oleic MUFA) was inversely correlated with CVD. However, it was shown that dietary oleic FA intake did not affect its plasma levels, since the concentrations of oleic MUFA in the blood are regulated by other factors, including *de novo* synthesis from stearic MUFA [47].

The results of the MESA (The Multi-Ethnic Study of Atherosclerosis, USA) study [47] show that elevated levels of oleic MUFA in plasma phospholipids may be a risk factor for the development of CVD and all-cause mortality. In the Aldo-DHF (Aldosterone in Diastolic Heart Failure, Germany) study [48], positive correlations were observed between the level of oleic MUFA and established cardiovascular risk factors, such as atherogenic dyslipidemia, dysglycemia, and obesity. In the population-based FINRISK study

(Finland) [49], it was determined that high levels of MUFA in the blood, including oleic FA, were associated with a higher risk of CVD. Similar results were obtained with respect to arterial hypertension [50] and inflammation [51]. Despite the relevance of studying the role/influence of MUFAs in the development of CVD and their risk factors, additional studies are needed on the influence of non-dietary factors, such as genetics or younger populations.

POLYUNSATURATED FATTY ACIDS

Recently, special attention has been paid to the role and importance of nutrients, especially long-chain omega-3 and omega-6 PUFAs. It has been shown that omega-3 PUFAs may be beneficial in various diseases and conditions, such as atherosclerosis [52], obesity [53], and inflammation [54]. However, the cardioprotective properties of omega-3 PUFAs are considered to be the most studied. The biological effects of omega-6 PUFAs are still poorly understood and are the subject of active debate [55]. Although most studies report that some omega-6 PUFAs are associated with a lower risk of CVD [56], they have powerful vasodilatory, antiplatelet, and antiarrhythmic effects [57].

The alpha-linolenic acid (C18:3, omega-3) is the most common omega-3 PUFA that can be obtained only from food (mainly from plant sources: flaxseed oil, walnuts, soy, etc.) [58]. One of the large metaanalyses of the Cochrane Database [59], which included 86 randomized controlled trials lasting at least 12 months, assessed the effect of increased omega-3 FA intake on overall mortality, CVD, obesity, and lipid profile. The results showed that an increase in alpha-linolenic PUFA slightly reduced the risk of cardiovascular events. A subsequent meta-analysis [60] including the results of 47 studies confirmed that increasing alpha-linolenic PUFA intake by 1 g / day was associated with reductions in triglycerides, total cholesterol, and LDL-C, thereby preventing CVD.

In a meta-analysis of 27 observational studies [61], data on the association of alpha-linolenic PUFA and the risk of developing CVD were summarized. Observations show that total exposure to C18:3 omega-3 PUFA is associated with a moderately lower risk of CVD. Within the PREDIMED study [62], it was found that in people with high cardiovascular risk, but without previous CVD, the alpha- PUFA intake was inversely correlated with all-cause mortality. The Alpha – Omega study [63] revealed a trend toward

a reduction in the risk of CVD with alpha-linolenic PUFA consumption in patients receiving modern cardiac treatment.

In a study of the relationship between the levels of alpha-linolenic PUFA in plasma and the risk of acute coronary syndrome, T.A. Zelniker et al. [64] found significant inverse associations of C18:3 omega-3 with a lower risk of sudden cardiac death, independent of traditional risk factors and lipid levels. And in a study on mice, it was shown that a diet rich in C18:3 omega-3 can protect against endothelial dysfunction and prevent the development of atherosclerosis by suppressing the inflammatory response and the formation of foam cells [65].

Eicosapentaenoic acid (C20:5, omega-3) is considered to be an essential omega-3 PUFA. It is found primarily in fish and other seafood but can be biosynthesized in small amounts from its main precursor, alpha-linolenic PUFA [66]. There is strong evidence that eicosapentaenoic PUFA has beneficial effects on endothelial function and increases the synthesis of eicosanoids (which dilate blood vessels and reduce thrombus formation and inflammation) [67]. In addition, its potential therapeutic effects on the atherosclerotic plaque include anti-inflammatory and antioxidant activity, reduction of macrophage and foam cell accumulation in lipid spots, reduction of monocyte adhesion, and increase in the thickness of the fibrous cap of the plaque [67–70].

The JELIS (Japan Eicosapentaenoic acid Lipid Intervention Study, Japan) study [71] showed that the introduction of eicosapentaenoic PUFA at a dose of 1.8 g / day led to a decrease in CVD by 19% in patients receiving statins and a decrease in LDL-C concentration in the blood by 25% after treatment. The results of the multicenter, randomized REDUCE-IT (Reduction of Cardiovascular Events with Icosapent Ethyl–Intervention Trial, USA) study [72] indicate that in patients with elevated triglyceride levels who received icosapent ethyl 4 g / day, the risk of major ischemic events, including sudden cardiac death, was significantly lower.

The OCEAN (Omacor Carotid Endarterectomy Intervention, UK) study [73] noted that higher levels of eicosapentaenoic PUFA in atherosclerotic plaques were associated with a decrease in the number of foam and T cells, less pronounced inflammation, and increased stability. Accordingly, the use of C20:5 omega-3 in individuals with a high risk of developing CVD as additional drug therapy helps reduce this risk [74].

Docosahexaenoic acid (C22:6, omega-3) is a very long-chain omega-3 PUFA found in high concentrations in fish, fish oil, and some algae [75]. Clinical studies using dietary supplements with high levels of docosahexaenoic PUFA have shown stable anti-inflammatory, antioxidant, antiatherogenic, and antiproliferative effects [76, 77]. In a double-blind, multigroup, placebo-controlled, randomized study [78], it was shown that C 22:6 omega-3 was more effective than C 20:5 omega-3 in reducing blood triglyceride levels, partly due to differential regulation of liver enzymes associated with lipogenesis. However, consumption of docosahexaenoic PUFA at a dose of ~3 g/day for 10 weeks may be more effective in reducing inflammatory markers, such as interleukin-18, tumor necrosis factor α, and C-reactive protein [79]. There is also evidence that consumption of docosahexaenoic PUFA increases not only C22:6 omega-3 in blood and tissues, but also C20:5 omega-3 eicosapentaenoic PUFA [80]. Moreover, the increase in the omega-3 FA index is significantly higher after supplementation with docosahexaenoic PUFA (2.7 g/day)[81]. Finally, a number of authors have found that docosahexaenoic PUFA causes a greater decrease in blood pressure, heart rate, and total peripheral resistance compared to eicosapentaenoic PUFA [82-84]. Thus, relatively high levels of free omega-3 PUFA may not always be associated with protection of the acutely damaged heart, but nevertheless have a beneficial effect on the body as a whole.

Linoleic acid (C18:2, omega-6) is the main dietary source of other omega-6 PUFAs, such as gamma-linolenic acid, dihomo-gamma-linolenic acid, and arachidonic acid. Linoleic acid is mainly obtained from vegetable oils [85]. There is increasing evidence that high linoleic acid levels are significantly associated with a reduction in the risk of development and mortality from CVD [86, 87]. According to the results of the Cochrane Database meta-analysis [88], which included 19 randomized controlled trials, higher intake of linoleic PUFA instead of SFA or carbohydrates reduced the risk of developing MI and total serum cholesterol by 6%. According to a metaanalysis of 30 prospective studies from 13 countries [56], higher levels of linoleic PUFA in vivo were associated with a lower risk of CVD, in particular, mortality from stroke.

In a meta-analysis of observational studies [86], high serum/dietary omega-6 C18:2 levels were inversely proportional to the risk of hypertension. In addition, the results of the International Study of

Macro-Micronutrients and Blood Pressure Study (INTERMAP) [89] show that dietary linoleic PUFA intake may contribute to the prevention and control of unfavorable blood pressure levels in the general population. In a study aimed at investigating the risks of CVD in communities (CIRCS) [21], it was found that serum levels of omega-6 linoleic PUFA were inversely associated with the risk of CHD.

In a Mendelian randomization study [90], it was shown that higher serum omega-6 C18:2 levels were inversely associated with lower levels of lipids, including LDL-C, HDL-C, and total cholesterol. In general, it can be noted that enriching the diet with a moderate amount of linoleic acid-rich oil may reduce the risk of cardiometabolic diseases [91].

Dihomo-gamma-linolenic acid (C20:3, omega-6) is considered to be one of the key omega-6 PUFAs, which has antiatherogenic effects. It inhibits the formation of foam cells, reduces the proliferation of endothelial cells, improves mitochondrial function, etc. [92]. By means of enzymatic activity, gammalinolenic acid (C18:3, omega-6) is very quickly converted into dihomo-gamma-linolenic PUFA. The latter, in turn, can be metabolized into the antiinflammatory eicosanoid - prostaglandin E1, via the cyclooxygenase pathway [93]. In the body, it is found in lipids (primarily phospholipids) and most cells, and C20:3 omega-6 levels are consistently increased following C18:3 omega-6 supplementation [94]. In mice, dihomo-gamma-linolenic PUFA supplementation has been shown to reduce aortic lipid content, along with macrophage and smooth muscle cell levels and ICAM-1 and VCAM-1 expression [93].

Few studies have shown an association between low levels of dihomo-gamma-linolenic PUFA and the severity of CHD [95]. In the OMEMI study [96], low serum levels of dihomo-gamma-linolenic PUFA were associated with an increased risk of all-cause mortality in elderly patients who had recently experienced MI. Similar results were obtained by S. Ouchi et al. [97], where the authors concluded that low levels of dihomogamma-linolenic PUFA in serum may be a predictor of permanent CVD (acute coronary syndrome, MI). In the work by T. Nagai et al. [98], lower levels of omega-6 PUFA, in particular C20:3, were associated with higher incidence of adverse events (death from all causes and observation of heart failure) after acute decompensated heart failure.

Finally, arachidonic acid (C20:4, omega-6), also known as eicosatetraenoic PUFA of the omega-6 class, is worth noting. It can enter the human body as part

of various foods (meat, eggs, salmon, vegetable oils, walnuts) or be formed by endogenous synthesis due to release from phospholipids in the cell membrane by cytosolic phospholipiase A2 (PLA2) [99]. It is usually esterified as triglycerides or glycerophospholipids to maintain cell membrane structure and function. It is well known that arachidonic PUFA can compete with omega-3 eicosapentaenoic PUFA for cyclooxygenase and lipoxygenase *in vivo* [100]. The arachidonic PUFA and its metabolites play an important role in the functioning of the cardiovascular system. They act as vasodilators or vasoconstrictors and modulate vasodilation in pathological and physiological conditions [101].

Nevertheless, the results of studies on the associations of circulating or tissue levels of arachidonic PUFA with CVD are rather inconclusive. A meta-analysis of 30 prospective studies [56] did not support adverse cardiovascular effects of arachidonic PUFA. Moreover, the authors suggested that higher plasma C20:4 levels may be associated with a lower risk of developing CVD. In two population-based cohort studies conducted in the Netherlands [102], no association was found between arachidonic PUFA levels and the risk of developing CHD. In the analysis of data obtained from a retrospective registry of patients with acute hypertensive stroke [103], lower serum arachidonic PUFA levels were independently associated with poor functional outcome in acute intracerebral hemorrhage. According to the results of a study using genetic variants [104], positive associations of arachidonic PUFA with atherosclerotic CVD and venous thromboembolism were found. When studying the content of arachidonic PUFA in adipose tissue, a positive association with the risk of MI in the Danish Prospective Cohort Study (DCH) was established [105].

CONCLUSION

Thus, the study of the influence of FA on the development of CVD is a promising area of research. Data on the associations of different SFA, MUFA, and PUFA with lipid and lipoprotein parameters and inflammatory markers of CVD may be of interest for obtaining new data clarifying and supplementing the mechanisms of the effect of FA on the cardiovascular system.

REFERENCES

 Boytsov S.A., Drapkina O.M., Shlyakhto E.V., Konradi A.O., Balanova Yu.A., Zhernakova Yu.V. et al. Epidemiology of Cardiovascular Diseases and their Risk Factors in Regions of

- the Russian Federation (ESSE-RF Study). Ten Years Later. *Cardiovascular Therapy and Prevention*. 2021;20(5):3007. (In Russ.). DOI: 10.15829/1728-8800-2021-3007.
- Latfullin I.A. Coronary Heart Disease: the Main Risk Factors, Treatment. Kazan: Kazan (Volga Region) Federal University, 2017:426 (In Russ.).
- 3. Badeinikova K.K., Mamedov M.N. Early Markers of Atherosclerosis: Predictors of Cardiovascular Events. *Profilakticheskaya Meditsina Preventive Medicine*. 2023;26(1):103–108. (In Russ.). DOI:10.17116/profmed202326011103.
- Sergienko I.V., Ansheles A.A. Pathogenesis, Diagnosis and Treatment of Atherosclerosis: Practical Aspects. *Russian Car-diology Bulletin*. 2021;16(1):64–72. (In Russ.). DOI:10.17116/ Cardiobulletin20211601164.
- 5. The top-10 causes of death in the world (fact sheet). World Health Organization., 2020. https://www.who.int/ru/news-room/fact-sheets/detail/the-top-10-causes-of-death
- Kotlyarov S., Kotlyarova A. Involvement of Fatty Acids and Their Metabolites in the Development of Inflammation in Atherosclerosis. *Int. J. Mol. Sci.* 2022;23(3):1308. DOI: 10.3390/ijms23031308.
- Chen X., Liu L., Palacios G., Gao J., Zhang N., Li G. et al. Plasma metabolomics reveals biomarkers of the atherosclerosis. *J. Sep. Sci.* 2010;33(17-18):2776–2783. DOI: 10.1002/ jssc.201000395.
- 8. Kotlyarov S., Kotlyarova A. Clinical significance of polyunsaturated fatty acids in the prevention of cardiovascular diseases. *Front. Nutr.* 2022;9:998291. DOI: 10.3389/fnut.2022.998291.
- 9. Ghosh A., Gao L., Thakur A., Siu P.M., Lai C.W.K. Role of free fatty acids in endothelial dysfunction. *J. Biomed. Sci.* 2017;24(1):50. DOI: 10.1186/s12929-017-0357-5.
- Zotov V.A., Bessonov V.V., Risnik D.V. Methodological Aspects of the Analysis of Fatty Acids in Biological Samples. Applied Biochemistry and Microbiology. 2022;58(1):90–104. (In Russ.). DOI 10.31857/S0555109922010111.
- Gizinger O.A. The Role of Short- and Medium-Chain Fatty Acids in Homeostatic Regulation Reactions. *Therapist*. 2021;9:45–51. (In Russ.). DOI: 10.33920/MED-12-2109-05.
- Sacks F.M., Lichtenstein A.H., Wu J.H.Y., Appel L.J., Creager M.A., Kris-Etherton P.M. et al. Dietary fats and cardiovascular disease: a presidential advisory from the American Heart Association. *Circulation*. 2017;136(3):e1–e23. DOI: 10.1161/CIR.0000000000000010.
- Annevelink C.E., Sapp P.A., Petersen K.S., Shearer G.C., Kris-Etherton P.M. Diet-derived and diet-related endogenously produced palmitic acid: Effects on metabolic regulation and cardiovascular disease risk. *J. Clin. Lipidol*. 2023;17(5):577– 586. DOI: 10.1016/j.jacl.2023.07.005.
- Carta G., Murru E., Banni S., Manca C. Palmitic acid: physiological role, metabolism and nutritional implications. *Front. Physiol.* 2017;8:902. DOI: 10.3389/fphys.2017.00902.
- Fatima S., Hu X., Gong R.H., Huang C., Chen M., Wong H.L.X. et al. Palmitic acid is an intracellular signaling molecule involved in disease development. *Cell Mol. Life Sci.* 2019;76(13):2547–2557. DOI: 10.1007/s00018-019-03092-7.
- Innis S.M. Fatty acids and early human development. *Early Hum. Dev.* 2007;83(12):761–766. DOI: 10.1016/j.earlhumdev.2007.09.004.

- 17. Titov V. N., Aripovsky A.V., Kaba S. I., Kolesnik P. O., Vejdel M. I., Shiryaeva Yu. K. The Individual Fatty Acids in Blood Plasma, Erythrocytes, and Lipoproteins. The Comparison of Test Results of Patients with Coronary Heart Disease and Volunteers. *Clinical Laboratory Diagnostics*. 2012;7:3–8. (In Russ.).
- 18. Domínguez-López I., Arancibia-Riveros C., Casas R., Tresserra-Rimbau A., Razquin C., Martínez-González M.Á. et al. Changes in plasma total saturated fatty acids and palmitic acid are related to pro-inflammatory molecule IL-6 concentrations after nutritional intervention for one year. *Biomed. Pharmacother*. 2022;150:113028. DOI: 10.1016/j.biopha.2022.113028.
- Kleber M.E., Delgado G.E., Dawczynski C., Lorkowski S., März W, von Schacky C. Saturated fatty acids and mortality in patients referred for coronary angiography-The Ludwigshafen Risk and Cardiovascular Health study. *J. Clin. Lipidol*. 2018;12(2):455–463.e3. DOI: 10.1016/j.jacl.2018.01.007.
- Lee Y., Lai H.T.M., de Oliveira Otto M.C., Lemaitre R.N., McKnight B., King I.B. et al. Serial Biomarkers of De Novo Lipogenesis Fatty Acids and Incident Heart Failure in Older Adults: The Cardiovascular Health Study. *J. Am. Heart Assoc*. 2020;9(4):e014119. DOI: 10.1161/JAHA.119.014119.
- Chei C.L., Yamagishi K., Kitamura A., Kiyama M., Sankai T., Okada T. et al. Serum Fatty Acid and Risk of Coronary Artery Disease - Circulatory Risk in Communities Study (CIRCS). *Circ. J.* 2018;82(12):3013–3020. DOI: 10.1253/circj.CJ-18-0240.
- Mensink R.P. Effects of saturated fatty acids on serum lipids and lipoproteins: a systematic review and regression analysis. Geneva: World Health Organization, 2016:72.
- 23. Praagman J., de Jonge E.A., Kiefte-de Jong J.C., Beulens J.W., Sluijs I., Schoufour J.D. et al. Dietary saturated fatty acids and coronary heart disease risk in a dutch middle-aged and elderly population. *Arterioscler. Thromb. Vasc. Biol.* 2016;36(9):2011–2018. DOI: 10.1161/ATVBA-HA.116.307578.
- 24. Zong G., Li Y., Wanders A.J., Alssema M., Zock P.L., Willett W.C. et al. Intake of individual saturated fatty acids and risk of coronary heart disease in US men and women: two prospective longitudinal cohort studies. *BMJ*. 2016;355:i5796. DOI: 10.1136/bmj.i5796.
- Yuan S., Bäck M., Bruzelius M., Mason A.M., Burgess S., Larsson S. Plasma Phospholipid Fatty Acids, FADS1 and Risk of 15 Cardiovascular Diseases: A Mendelian Randomisation Study. *Nutrients*. 2019;11(12):3001. DOI: 10.3390/ nu11123001.
- Khaw K.T., Friesen M.D., Riboli E., Luben R., Wareham N. Plasma phospholipid fatty acid concentration and incident coronary heart disease in men and women: the EPIC-Norfolk prospective study. *PLoS Med.* 2012;9(7):e1001255. DOI: 10.1371/journal.pmed.1001255.
- 27. Lai H.T.M., de Oliveira Otto M.C., Lee Y., Wu J.H.Y., Song X., King I.B. et al. Serial Plasma Phospholipid Fatty Acids in the De Novo Lipogenesis Pathway and Total Mortality, Cause-Specific Mortality, and Cardiovascular Diseases in the Cardiovascular Health Study. *J. Am. Heart Assoc*. 2019;8(22):e012881. DOI: 10.1161/JAHA.119.012881.
- Van Rooijen M.A., Plat J., Blom W.A.M., Zock P.L., Mensink R.P. Dietary stearic acid and palmitic acid do not differ-

- ently affect ABCA1-mediated cholesterol efflux capacity in healthy men and postmenopausal women: A randomized controlled trial. *Clin. Nutr.* 2021;40(3):804-811. DOI: 10.1016/j. clnu.2020.08.016.
- 29. Praagman J., Beulens J.W., Alssema M., Zock P.L., Wanders A.J., Sluijs I. et al. The association between dietary saturated fatty acids and ischemic heart disease depends on the type and source of fatty acid in the European Prospective Investigation into Cancer and Nutrition-Netherlands cohort. Am. J. Clin. Nutr. 2016;103(2):356–365. DOI: 10.3945/ajcn.115.122671.
- Zazula R., Moravec M., Pehal F., Nejtek T., Protuš M., Müller M. Myristic acid serum levels and their significance for diagnosis of systemic inflammatory response, sepsis, and bacteraemia. *J. Pers. Med.* 2021;11(4):306. DOI: 10.3390/ jpm11040306.
- 31. Rioux V., Catheline D., Legrand P. In rat hepatocytes, myristic acid occurs through lipogenesis, palmitic acid shortening and lauric acid elongation. *Animal*. 2007;1(6):820–826. DOI: 10.1017/S1751731107000122.
- 32. Noto D., Fayer F., Cefalù A.B., Altieri I., Palesano O., Spina R. et al. Myristic acid is associated to low plasma HDL cholester-ol levels in a Mediterranean population and increases HDL catabolism by enhancing HDL particles trapping to cell surface proteoglycans in a liver hepatoma cell model. *Atherosclerosis*. 2016;246:50–56. DOI: 10.1016/j.atherosclerosis.2015.12.036.
- Ebbesson S.O., Voruganti V.S., Higgins P.B., Fabsitz R.R., Ebbesson L.O., Laston S. et al. Fatty acids linked to cardiovascular mortality are associated with risk factors. *Int. J. Circumpolar. Health.* 2015;74:28055. DOI: 10.3402/ijch.v74.28055.
- Hooper L., Martin N., Jimoh O.F., Kirk C., Foster E., Abdelhamid A.S. Reduction in saturated fat intake for cardiovascular disease. *Cochrane Database Syst. Rev.* 2020;8(8):CD011737. DOI: 10.1002/14651858.CD011737.pub3.
- 35. Legrand-Poels S., Esser N., L'homme L., Scheen A., Paquot N., Piette J. Free fatty acids as modulators of the NLRP3 inflammasome in obesity/type 2 diabetes. *Biochem. Pharmacol.* 2014;92(1):131–141. DOI: 10.1016/j.bcp.2014.08.013.
- 36. Frigolet M.E., Gutiérrez-Aguilar R. The role of the novel lipokine palmitoleic acid in health and disease. *Adv. Nutr.* 2017;8(1):173S–181S. DOI: 10.3945/an.115.011130.
- Rocha D.M., Bressan J., Hermsdorff H.H. The role of dietary fatty acid intake in inflammatory gene expression: a critical review. *Sao Paulo Med. J.* 2017;135(2):157–168. DOI: 10.1590/1516-3180.2016.008607072016.
- Takenouchi Y., Seki Y., Shiba S., Ohtake K., Nobe K., Kasono K. Effects of dietary palmitoleic acid on vascular function in aorta of diabetic mice. *BMC Endocr. Disord*. 2022;22(1):103. DOI: 10.1186/s12902-022-01018-2.
- Cao H., Gerhold K., Mayers J.R., Wiest M.M., Watkins S.M., Hotamisligil G.S. Identification of a lipokine, a lipid hormone linking adipose tissue to systemic metabolism. *Cell*. 2008;134(6):933–944. DOI: 10.1016/j.cell.2008.07.048.
- Guo X., Li H., Xu H., Halim V., Zhang W., Wang H. et al. Palmitoleate induces hepatic steatosis but suppresses liver inflammatory response in mice. *PLoS One*. 2012;7(6):e39286. DOI: 10.1371/journal.pone.0039286.
- 41. Mozaffarian D., de Oliveira Otto M.C., Lemaitre R.N., Fretts A.M., Hotamisligil G., Tsai M.Y. et al. trans-Palmitoleic acid, oth-

- er dairy fat biomarkers, and incident diabetes: the Multi-Ethnic Study of Atherosclerosis (MESA). *Am. J. Clin. Nutr.* 2013;97(4):854–861. DOI: 10.3945/ajcn.112.045468.
- Luan D., Wang D., Campos H., Baylin A. Adipose tissue palmitoleic acid is inversely associated with nonfatal acute myocardial infarction in Costa Rican adults. *Nutr. Metab. Cardiovasc. Dis.* 2018;28(10):973–979. DOI: 10.1016/j.numecd.2018.05.004.
- 43. De Souza C.O., Vannice G.K., Rosa Neto J.C., Calder P.C. Is palmitoleic acid a plausible nonpharmacological strategy to prevent or control chronic metabolic and inflammatory disorders? *Mol. Nutr. Food Res.* 2018;62(1). DOI: 10.1002/ mnfr.201700504.
- 44. Chan K.L., Pillon N.J., Sivaloganathan D.M., Costford S.R., Liu Z., Théret M. et al. Palmitoleate reverses high fat-induced proinflammatory macrophage polarization via AMP-activated protein kinase (AMPK). *J. Biol. Chem.* 2015;290(27):16979– 16988. DOI: 10.1074/jbc.M115.646992.
- 45. Souza C.O., Teixeira A.A., Biondo L.A., Silveira L.S., Calder P.C., Rosa Neto J.C. Palmitoleic acid reduces the inflammation in LPS-stimulated macrophages by inhibition of NFκB, independently of PPARs. *Clin. Exp. Pharmacol. Physiol.* 2017;44(5):566–575. DOI: 10.1111/1440-1681.
- 46. Guasch-Ferré M., Hu F.B., Martínez-González M.A., Fitó M., Bulló M., Estruch R. et al. Olive oil intake and risk of cardiovascular disease and mortality in the PREDIMED Study. *BMC Med.* 2014;12:78. DOI: 10.1186/1741-7015-12-78.
- 47. Steffen B.T., Duprez D., Szklo M., Guan W., Tsai M.Y. Circulating oleic acid levels are related to greater risks of cardiovascular events and all-cause mortality: The Multi-Ethnic Study of Atherosclerosis. *J. Clin. Lipidol.* 2018;12(6):1404–1412. DOI: 10.1016/j.jacl.2018.08.004.
- 48. Bock M., von Schacky C., Scherr J., Lorenz E., Lechner B., Krannich A. et al. De novo lipogenesis-related monounsaturated fatty acids in the blood are associated with cardiovascular risk factors in HFpEF patients. *J. Clin. Med.* 2023;12(15):4938. DOI: 10.3390/jcm12154938.
- Würtz P., Havulinna A.S., Soininen P., Tynkkynen T., Prieto-Merino D., Tillin T. et al. Metabolite profiling and cardio-vascular event risk: a prospective study of 3 population-based cohorts. *Circulation*. 2015;131(9):774–785. DOI: 10.1161/CIRCULATIONAHA.114.013116.
- Lin Y.T., Salihovic S., Fall T., Hammar U., Ingelsson E., Ärnlöv J. et al. Global plasma metabolomics to identify potential biomarkers of blood pressure progression. *Arterioscler. Thromb. Vasc. Biol.* 2020;40(8):e227–e237. DOI: 10.1161/ATVBAHA.120.314356.
- 51. Mika A., Sikorska-Wiśniewska M., Małgorzewicz S., Stepnowski P., Dębska-Ślizień A., Śledziński T. et al. Potential contribution of monounsaturated fatty acids to cardiovascular risk in chronic kidney disease. *Pol. Arch. Intern. Med.* 2018;128(12):755–763. DOI: 10.20452/pamw.4376.
- 52. Lands B. A critique of paradoxes in current advice on dietary lipids. *Prog. Lipid. Res.* 2008;47(2):77–106. DOI: 10.1016/j. plipres.2007.12.001.
- Alvheim A.R., Malde M.K., Osei-Hyiaman D., Lin Y.H., Pawlosky R.J., Madsen L. et al. Dietary linoleic acid elevates endogenous 2-AG and anandamide and induces obesity. *Obe-*

- sity (Silver Spring). 2012;20(10):1984–1894. DOI:10.1038/oby.2012.38.
- Maroon J.C., Bost J.W. Omega-3 fatty acids (fish oil) as an anti-inflammatory: an alternative to nonsteroidal anti-inflammatory drugs for discogenic pain. *Surg. Neurol.* 2006;65(4):326–331. DOI: 10.1016/j.surneu.2005.10.023.
- 55. Garg P.K., Guan W., Nomura S., Weir N., Karger A.B., Duprez D. et al. Plasma ω-3 and ω-6 PUFA Concentrations and Risk of Atrial Fibrillation: The Multi-Ethnic Study of Atherosclerosis. J. Nutr. 2021;151(6):1479–1486. DOI: 10.1093/jn/nxab016.
- 56. Marklund M., Wu J.H.Y., Imamura F., Del Gobbo L.C., Fretts A., de Goede J. et al. Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) Fatty Acids and Outcomes Research Consortium (FORCE). Biomarkers of dietary omega-6 fatty acids and incident cardiovascular disease and mortality. *Circulation*. 2019;139(21):2422–2436. DOI: 10.1161/CIRCULATIONAHA.118.038908.
- 57. Das U.N. Essential fatty acids and their metabolites could function as endogenous HMG-CoA reductase and ACE enzyme inhibitors, anti-arrhythmic, anti-hypertensive, anti-atherosclerotic, anti-inflammatory, cytoprotective, and cardioprotective molecules. *Lipids Health Dis.* 2008;7:37. DOI: 10.1186/1476-511X-7-37.
- 58. Edel A.L., Patenaude A.F., Richard M.N., Dibrov E., Austria J.A., Aukema H.M. et al. The effect of flaxseed dose on circulating concentrations of alpha-linolenic acid and secoisolariciresinol diglucoside derived enterolignans in young, healthy adults. *Eur. J. Nutr.* 2016;55(2):651–663. DOI: 10.1007/s00394-015-0885-2.
- Abdelhamid A.S., Brown T.J., Brainard J.S., Biswas P., Thorpe G.C., Moore H.J. et al. Omega-3 fatty acids for the primary and secondary prevention of cardiovascular disease. *Cochrane Database Syst. Rev.* 2018;11(11):CD003177. DOI: 10.1002/14651858.CD003177.pub4.
- 60. Yue H., Qiu B., Jia M., Liu W., Guo X.F., Li N. et al. Effects of α-linolenic acid intake on blood lipid profiles: a systematic review and meta-analysis of randomized controlled trials. *Crit. Rev. Food Sci. Nutr.* 2021;61(17):2894–2910. DOI: 10.1080/10408398.2020.1790496.
- 61. Pan A., Chen M., Chowdhury R., Wu J.H., Sun Q., Campos H. et al. α-Linolenic acid and risk of cardiovascular disease: a systematic review and meta-analysis. *Am. J. Clin. Nutr.* 2012;96(6):1262–1273. DOI: 10.3945/ajcn.112.044040.
- 62. Sala-Vila A., Guasch-Ferré M., Hu F.B., Sánchez-Tainta A., Bulló M., Serra-Mir M. et al. Dietary α-Linolenic Acid, Marine ω-3 Fatty Acids, and Mortality in a Population With High Fish Consumption: Findings From the PREvención con DIeta MEDiterránea (PREDIMED) Study. *J. Am. Heart As*soc. 2016;5(1):e002543. DOI: 10.1161/JAHA.115.002543.
- Kromhout D., Giltay E.J., Geleijnse J.M.; Alpha Omega Trial Group. n-3 fatty acids and cardiovascular events after myocardial infarction. *N. Engl. J. Med.* 2010;363(21):2015–2026. DOI: 10.1056/NEJMoa1003603.
- 64. Zelniker T.A., Morrow D.A., Scirica B.M., Furtado J.D., Guo J., Mozaffarian D. et al. Plasma omega-3 fatty acids and the risk of cardiovasculareEvents in patients after an acute coronary syndrome in MERLIN-TIMI 36. *J. Am. Heart Assoc.* 2021;10(8):e017401. DOI: 10.1161/JAHA.120.017401.

- 65. Winnik S., Lohmann C., Richter E.K., Schäfer N., Song W.L., Leiber F. et al. Dietary α-linolenic acid diminishes experimental atherogenesis and restricts T cell-driven inflammation. *Eur. Heart J.* 2011;32(20):2573–2584. DOI: 10.1093/eurheartj/ehq501.
- Abedi E., Sahari M.A. Long-chain polyunsaturated fatty acid sources and evaluation of their nutritional and functional properties. *Food Sci. Nutr.* 2014;2(5):443–463. DOI: 10.1002/ fsn3.121.
- Borow K.M., Nelson J.R., Mason R.P. Biologic plausibility, cellular effects, and molecular mechanisms of eicosapentaenoic acid (EPA) in atherosclerosis. *Atherosclerosis*. 2015;242(1):357–366. DOI: 10.1016/j.atherosclerosis.2015.07.035.
- 68. Budoff M. Triglycerides and triglyceride-rich lipoproteins in the causal pathway of cardiovascular disease. *Am. J. Cardiol.* 2016;118(1):138–145. DOI: 10.1016/j.amjcard.2016.04.004.
- Gdula-Argasińska J., Czepiel J., Woźniakiewicz A., Wojtoń K., Grzywacz A., Woźniakiewicz M. et al. n-3 Fatty acids as resolvents of inflammation in the A549 cells. *Pharmacol. Rep.* 2015;67(3):610–615. DOI: 10.1016/j.pharep.2015.01.001.
- Nelson J.R., Wani O., May H.T., Budoff M. Potential benefits of eicosapentaenoic acid on atherosclerotic plaques. *Vascul. Pharmacol.* 2017;91:1–9. DOI: 10.1016/j.vph.2017.02.004.
- Itakura H., Yokoyama M., Matsuzaki M., Saito Y., Origasa H., Ishikawa Y. et al. Relationships between plasma fatty acid composition and coronary artery disease. *J. Atheroscler*. *Thromb*. 2011;18(2):99–107. DOI: 10.5551/jat.5876.
- 72. Bhatt D.L., Steg P.G., Miller M., Brinton E.A., Jacobson T.A., Ketchum S.B. et al. Cardiovascular risk reduction with icosapent ethyl for hypertriglyceridemia. *N. Engl. J. Med.* 2019;380(1):11–22. DOI: 10.1056/NEJMoa1812792.
- 73. Cawood A.L., Ding R., Napper F.L., Young R.H., Williams J.A., Ward M.J. et al. Eicosapentaenoic acid (EPA) from highly concentrated n-3 fatty acid ethyl esters is incorporated into advanced atherosclerotic plaques and higher plaque EPA is associated with decreased plaque inflammation and increased stability. *Atherosclerosis*. 2010;212(1):252–259. DOI: 10.1016/j. atherosclerosis.2010.05.022.
- Singh S., Arora R.R., Singh M., Khosla S. Eicosapentaenoic acid versus docosahexaenoic acid as options for vascular risk prevention: a fish story. *Am. J. Ther.* 2016;23(3):e905–910. DOI: 10.1097/MJT.0000000000000165.
- Honda K.L., Lamon-Fava S., Matthan N.R., Wu D., Lichtenstein A.H. Docosahexaenoic acid differentially affects TNFα and IL-6 expression in LPS-stimulated RAW 264.7 murine macrophages. *Prostaglandins Leukot. Essent. Fatty Acids*. 2015;97:27–34. DOI: 10.1016/j.plefa.2015.03.002.
- Peris-Martínez C., Piá-Ludeña J.V., Rog-Revert M.J., Fernández-López E., Domingo J.C. Antioxidant and Anti-Inflammatory Effects of Oral Supplementation with a Highly-Concentrated Docosahexaenoic Acid (DHA) Triglyceride in Patients with Keratoconus: A Randomized Controlled Preliminary Study. *Nutrients*. 2023;15(5):1300. DOI: 10.3390/nu15051300.
- 77. So J., Wu D., Lichtenstein A.H., Tai A.K., Matthan N.R., Maddipati K.R. et al. EPA and DHA differentially modulate monocyte inflammatory response in subjects with chronic inflammation in part via plasma specialized pro-resolving lipid mediators: A randomized, double-blind, crossover study. Ath-

- *erosclerosis*. 2021;316:90–98. DOI: 10.1016/j.atherosclerosis.2020.11.018.
- 78. Klingel S.L., Metherel A.H., Irfan M., Rajna A., Chabowski A., Bazinet R.P. et al. EPA and DHA have divergent effects on serum triglycerides and lipogenesis, but similar effects on lipoprotein lipase activity: a randomized controlled trial. *Am. J. Clin. Nutr.* 2019;110(6):1502–1509. DOI: 10.1093/ajcn/nqz234.
- 79. Allaire J., Couture P., Leclerc M., Charest A., Marin J., Lépine M.C. et al. A randomized, crossover, head-to-head comparison of eicosapentaenoic acid and docosahexaenoic acid supplementation to reduce inflammation markers in men and women: the Comparing EPA to DHA (ComparED) Study. Am. J. Clin. Nutr. 2016;104(2):280–287. DOI: 10.3945/ajcn.116.131896.
- 80. Metherel A.H., Irfan M., Klingel S.L., Mutch D.M., Bazinet R.P. Compound-specific isotope analysis reveals no retroconversion of DHA to EPA but substantial conversion of EPA to DHA following supplementation: a randomized control trial. *Am. J. Clin. Nutr.* 2019;110(4):823–831. DOI: 10.1093/ajcn/nqz097.
- 81. Allaire J., Harris W.S., Vors C., Charest A., Marin J., Jackson K.H et al. Supplementation with high-dose docosahexaenoic acid increases the Omega-3 Index more than high-dose eicosapentaenoic acid. *Prostaglandins Leukot. Essent. Fatty Acids*. 2017;120:8–14. DOI: 10.1016/j.plefa.2017.03.008.
- 82. Lee J.B., Notay K., Klingel S.L., Chabowski A., Mutch D.M., Millar P.J. Docosahexaenoic acid reduces resting blood pressure but increases muscle sympathetic outflow compared with eicosapentaenoic acid in healthy men and women. *Am. J. Physiol. Heart Circ. Physiol.* 2019;316(4):H873–H881. DOI: 10.1152/ajpheart.00677.2018.
- 83. Rontoyanni V.G., Hall W.L., Pombo-Rodrigues S., Appleton A., Chung R., Sanders T.A. A comparison of the changes in cardiac output and systemic vascular resistance during exercise following high-fat meals containing DHA or EPA. *Br. J. Nutr.* 2012;108(3):492–499. DOI: 10.1017/S0007114511005721.
- 84. Rousseau-Ralliard D., Moreau D., Guilland J.C., Raederstorff D., Grynberg A. Docosahexaenoic acid, but not eicosapentaenoic acid, lowers ambulatory blood pressure and shortens interval QT in spontaneously hypertensive rats in vivo. *Prostaglandins Leukot. Essent. Fatty Acids.* 2009;80(5–6):269–277. DOI: 10.1016/j.plefa.2009.03.003.
- Choque B., Catheline D., Rioux V., Legrand P. Linoleic acid: between doubts and certainties. *Biochimie*. 2014;96:14–21. DOI: 10.1016/j.biochi.2013.07.012.
- 86. Hajihashemi P., Feizi A., Heidari Z., Haghighatdoost F. Association of omega-6 polyunsaturated fatty acids with blood pressure: A systematic review and meta-analysis of observational studies. *Crit. Rev. Food Sci. Nutr.* 2023;63(14):2247–2259. DOI: 10.1080/10408398.2021.1973364.
- 87. Harris W.S., Mozaffarian D., Rimm E., Kris-Etherton P., Rudel L.L., Appel L.J. et al. Omega-6 fatty acids and risk for cardiovascular disease: a science advisory from the American Heart Association Nutrition Subcommittee of the Council on Nutrition, Physical Activity, and Metabolism; Council on Cardiovascular Nursing; and Council on Epidemiology and Prevention. *Circulation*. 2009;119(6):902–907. DOI: 10.1161/CIRCULATIONAHA.108.191627.

- 88. Hooper L., Al-Khudairy L., Abdelhamid A.S., Rees K., Brainard J.S., Brown T.J. et al. Omega-6 fats for the primary and secondary prevention of cardiovascular disease. *Cochrane Database Syst. Rev.* 2018;7(7):CD011094. DOI: 10.1002/14651858.CD011094.pub3.
- Miura K., Stamler J., Nakagawa H., Elliott P., Ueshima H., Chan Q. et al. Relationship of dietary linoleic acid to blood pressure. The International Study of Macro-Micronutrients and Blood Pressure Study [corrected]. *Hypertension*. 2008;52(2):408–414. DOI: 10.1161/HYPERTENSIONAHA.108.112383.
- Zhao J.V., Schooling C.M. Effect of linoleic acid on ischemic heart disease and its risk factors: a Mendelian randomization study. *BMC Med.* 2019;17(1):61. DOI: 10.1186/s12916-019-1293-x.
- Cole R.M., Angelotti A., Sparagna G.C., Ni A., Belury M.A. Linoleic Acid-Rich Oil Alters Circulating Cardiolipin Species and Fatty Acid Composition in Adults: A Randomized Controlled Trial. *Mol. Nutr. Food Res.* 2022;66(15):e2101132. DOI: 10.1002/mnfr.202101132.
- 92. Gallagher H., Williams J.O., Ferekidis N., Ismail A., Chan Y.H., Michael D.R. et al. Dihomo-γ-linolenic acid inhibits several key cellular processes associated with atherosclerosis. *Biochim. Biophys. Acta Mol. Basis Dis.* 2019;1865(9):2538– 2550. DOI: 10.1016/j.bbadis.2019.06.011.
- Takai S., Jin D., Kawashima H., Kimura M., Shiraishi-Tateishi A., Tanaka T. et al. Anti-atherosclerotic effects of dihomo-gamma-linolenic acid in ApoE-deficient mice. *J. Atheroscler. Thromb.* 2009;16(4):480–489. DOI: 10.5551/jat.no430.
- Sergeant S., Rahbar E., Chilton F.H. Gamma-linolenic acid, Dihommo-gamma linolenic, Eicosanoids and Inflammatory Processes. *Eur. J. Pharmacol.* 2016;785:77–86. DOI: 10.1016/j.ejphar.2016.04.020.
- 95. Hadj Ahmed S., Kaoubaa N., Kharroubi W., Zarrouk A., Najjar M.F., Batbout F. et al. Association of plasma fatty acid alteration with the severity of coronary artery disease lesions in Tunisian patients. *Lipids Health Dis.* 2017;16(1):154. DOI: 10.1186/s12944-017-0538-y.
- 96. Nilsen D.W.T., Myhre P.L., Kalstad A., Schmidt E.B., Arnesen H., Seljeflot I. Serum levels of dihomo-gamma (γ)-linolenic acid (DGLA) are inversely associated with linoleic acid and total death in elderly patients with a recent myocardial infarction. *Nutrients*. 2021;13(10):3475. DOI: 10.3390/nu13103475.
- 97. Ouchi S., Miyazaki T., Shimada K., Sugita Y., Shimizu M., Murata A. et al. Decreased circulating dihomo-gamma-linolenic acid levels are associated with total mortality in patients with acute cardiovascular disease and acute decompensated heart failure. *Lipids Health Dis.* 2017;16(1):150. DOI: 10.1186/s12944-017-0542-2.
- 98. Nagai T., Honda Y., Sugano Y., Nishimura K., Nakai M., Honda S et al. Circulating omega-6, but not omega-3 polyunsaturated fatty acids, are associated with clinical autcomes in patients with acute decompensated heart failure. *PLoS One*. 2016;11(11):e0165841. DOI: 10.1371/journal. pone.0165841.
- Sonnweber T., Pizzini A., Nairz M., Weiss G., Tancevski I. Arachidonic acid metabolites in cardiovascular and metabolic diseases. *Int. J. Mol. Sci.* 2018;19(11):3285. DOI: 10.3390/ ijms19113285.

- 100. Zhang Y., Liu Y., Sun J., Zhang W., Guo Z., Ma Q. Arachidonic acid metabolism in health and disease. *Med. Comm.* (2020). 2023;4(5):e363. DOI: 10.1002/mco2.363.
- 101. Yang L., Mäki-Petäjä K., Cheriyan J., McEniery C., Wilkinson I.B. The role of epoxyeicosatrienoic acids in the cardiovascular system. *Br. J. Clin. Pharmacol.* 2015;80(1):28–44. DOI: 10.1111/bcp.12603.
- 102. De Goede J., Verschuren W.M., Boer J.M., Verberne L.D., Kromhout D., Geleijnse J.M. N-6 and N-3 fatty acid cholesteryl esters in relation to fatal CHD in a Dutch adult population: a nested case-control study and meta-analysis. *PLoS One*. 2013;8(5):e59408. DOI: 10.1371/journal.pone.0059408.
- 103. Takahashi J., Sakai K., Sato T., Takatsu H., Komatsu T., Mit-

- sumura H. et al. Serum arachidonic acid levels is a predictor of poor functional outcome in acute intracerebral hemorrhage. *Clin. Biochem.* 2021;98:42–47. DOI: 10.1016/j.clin-biochem.2021.09.012.
- 104. Zhang T., Zhao J.V., Schooling C.M. The associations of plasma phospholipid arachidonic acid with cardiovascular diseases: A Mendelian randomization study. *EBio Medicine*. 2021;63:103189. DOI: 10.1016/j.ebiom.2020.103189.
- 105. Nielsen M.S., Schmidt E.B., Stegger J., Gorst-Rasmussen A., Tjonneland A., Overvad K. Adipose tissue arachidonic acid content is associated with the risk of myocardial infarction: a Danish case-cohort study. *Atherosclerosis*. 2013;227(2):386–390. DOI: 10.1016/j.atherosclerosis.2012.12.035.

Author information

Shramko Viktoriya S. – Cand. Sc. (Medicine), Researcher, Laboratory of Clinical Biochemical and Hormonal Studies of Internal Diseases, Head of the Department of Clinical Biochemical and Molecular Genetic Research Methods, IIPM – Branch of IC&G SB RAS, Novosibirsk, Nosova@211.ru, https://orcid.org/0000-0002-0436-2549

Kashtanova Elena V. – Dr. Sc. (Biology), Associate Professor, Head of the Laboratory of Clinical Biochemical and Hormonal Studies of Internal Diseases, IIPM – Branch of IC&G SB RAS, Novosibirsk, elekastanova@yandex.ru, https://orcid.org/0000-0003-2268-4186

Stakhneva Ekaterina M. – Cand. Sc. (Biology), Senior Researcher, Laboratory of Clinical Biochemical and Hormonal Studies of Internal Diseases, IIPM –Branch of IC&G SB RAS, Novosibirsk, stahneva@yandex.ru, https://orcid.org/0000-0003-0484-6540;

Polonskaya Yana V. – Dr. Sc. (Biology), Senior Researcher, Laboratory of Clinical Biochemical and Hormonal Studies of Internal Diseases, IIPM –Branch of IC&G SB RAS, Novosibirsk, yana-polonskaya@yandex.ru, https://orcid.org/0000-0002-3538-0280

Ragino Yulia I. – Dr. Sc. (Biology), Professor, Corresponding Member of the Russian Academy of Sciences, Head of the IIPM – Branch of IC&G SB RAS, Chief Researcher at the Laboratory of Clinical Biochemical and Hormonal Studies of Internal Diseases; IIPM – Branch of IC&G SB RAS, Novosibirsk, ragino@mail.ru, https://orcid.org/0000-0002-4936-8362

(⊠) Shramko Viktoriya S., Nosova@211.ru

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